



Review

Cerebral Inflammation in X-Linked Adrenoleukodystrophy

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Abstract. X-linked adrenoleukodystrophy (X-ALD) is an inherited neurodegenerative disease that affects approximately 1 in 25 000 males. It is characterized by elevated levels of saturated very long chain fatty acids (VLCFA), i.e., >C22:0, particularly in ganglioside and cholesterol ester fractions of brain white matter and adrenal cortex. Failure of peroxisomal very long chain fatty acyl-CoA synthetase (VLCS) to activate these VLCFA prevents their degradation by peroxisomal β -oxidation. X-ALD maps to Xq28³³ and the gene encodes a peroxisomal membrane protein^{39, 40, 65} and not the gene for VLCS. The two most common forms of X-ALD are the cerebral (CER) form, with an inflammatory demyelinating reaction that resembles multiple sclerosis (MS), and adrenomyeloneuropathy (AMN), which involves the spinal cord and in which the inflammatory reaction is mild or absent. Investigations into the nature of the cerebral inflammatory demyelinating reaction in X-ALD will be the subject of this review.

Key words: X-linked adrenoleukodystrophy; inflammation; demyelination; cytokines; human leukocyte antigens.

Phenotypic Variability of X-ALD

The phenotypic presentation of X-ALD mutations is remarkably variable. Six phenotypes have been delineated in X-ALD hemizygotes that vary with respect to age of disease onset, the duration from age of onset to death, and the site of initial pathology.

The childhood cerebral form (CCER), in which there is a cerebral perivascular inflammatory demyelinating reaction, is the most common and severe form of X-ALD, occurring in 48% of approximately 1500 patients studied at the Kennedy Krieger Institute³⁸. The mean age of onset of neurological symptoms in CCER is 7.1 ± 1.7 years. In this illness, behavioral and cognitive deterioration progresses rapidly and patients typically die within 2 to 4 years of onset. In a small number of cerebral (CER) patients, cerebral involvement

begins in adolescence (5%) or adulthood (3%) but is otherwise similar to CCER in respect to its clinical manifestations, rates of progression and the inflammatory response. AMN, with initial and primary involvement of the spinal cord and peripheral nerves, is the second most common form, occurring in 25% of patients in the Kennedy Krieger Institute series. AMN manifests with slowly progressive paraparesis and sphincter disturbances, involves long tracts in the spinal cord mainly, and appears to be a distal axonopathy with secondary demyelination. The inflammatory response is mild or absent. The mean age of onset in AMN is 27 years, but can occur as late as >60 years. Progression is slow and patients live well into adulthood. There is some cerebral involvement (as evidenced by magnetic resonance imaging data) in approximately 40% of AMN patients during the later stages of the disease²⁵.

Abbreviations used: X-ALD – X-linked adrenoleukodystrophy, AMN – adrenomyeloneuropathy, CCER – childhood cerebral form of X-linked adrenoleukodystrophy, CER – cerebral form of X-linked adrenoleukodystrophy.

The designation Addison-only (AD) is applied to patients with the biochemical defect of X-ALD who have primary adrenal insufficiency but are free of demonstrable neurological involvement. Some individuals with X-ALD mutations and elevated VLCFA have no clinical manifestations. Although late onset cannot be ruled out, asymptomatic individuals in their 60s have been observed.

In the other inherited lipidoses (e.g. metachromatic leukodystrophy, globoid cell leukodystrophy, Niemann-Pick disease, GM2 gangliosidosis) there is a correlation between the extent of biochemical abnormality and clinical phenotype. In X-ALD, however, no correlation between phenotype and VLCFA level (the biochemical abnormality) in plasma, fibroblasts or red blood cells, or fibroblast VLCFA β -oxidation has been reported in a large number of patients^{1, 4, 36}. ANTOKU et al.¹ reported a difference in the mononuclear cell VLCFA ratio between 4 patients with childhood-adolescent CER and 4 patients with adult X-ALD (2 with AMN and 2 with adult CER).

It has been shown repeatedly that the same mutations in the X-ALD gene are associated with each of the six phenotypes^{2, 21, 24, 39}. Thus, there is no correlation between genotype at the X-linked locus and phenotype. In addition, all phenotypes segregate in the same X-ALD kindreds and 50% of kindreds have both CCER and AMN³⁸. This suggests that a factor other than the X-linked locus is involved in determining phenotype. Genetic segregation analysis of a large number of pedigrees and sib pair analysis independently suggest that an autosomal modifying gene determines whether or not individuals with X-ALD mutations develop cerebral inflammatory demyelinating lesions during childhood^{37, 58}.

The Cerebral Inflammatory Reaction in X-ALD

The pathogenetic mechanism of the cerebral inflammatory demyelinating reaction in X-ALD and the role of the protein product (ALDP) of the X-ALD gene at Xq28 in destructive demyelination are unknown. The inflammatory reaction in CER could be to an as yet unknown antigen (e.g. the accumulation of VLCFA or compounds containing VLCFA in the brain) with myelin-breakdown secondary to this inflammatory reaction. The mutated X-ALD protein might be antigenic but, as 69% of X-ALD patients (both CER and AMN) lack ALDP^{9, 22, 65}, it is clearly not required to initiate the cerebral inflammatory demyelinating reaction. Alternatively, elevated levels of VLCFA in myelin lipids in CER patients could lead to myelin instability and subsequent myelinolysis followed by an inflammatory reaction. Whatever the initiating event in the cerebral

inflammatory demyelinating reaction in CER, it has been hypothesized that myelin breakdown products initiate a cytokine cascade which may lead to breakdown of the blood brain barrier followed by a cellular-based reaction to a central nervous system (CNS)-specific antigen (since there is no inflammatory reaction in the peripheral nervous system (PNS))⁴⁷.

The inflammatory nature of the cerebral demyelination seen in X-ALD sets it apart from the demyelination seen in the other inheritable leukodystrophies (e.g. metachromatic leukodystrophy, globoid cell leukodystrophy) and aligns it more with MS, the most common immune-mediated cerebral inflammatory demyelinating disease. POWERS et al.⁴⁸ showed that in CER inflammatory demyelinating lesions, reactive astrocytes, macrophages and T cells are the most prevalent cellular elements, with very few B cells present. They showed that in CER the inflammatory infiltrates are located behind the active lesion edge and, unexpectedly, in subsequent secondary tract degeneration. In contrast, the inflammatory infiltrates in MS are found at the active demyelinating lesion edge⁵³. GRIFFIN et al.¹² identified both CD4⁺ and CD8⁺ T cells in CER lesions and also found a large number of B cells in perivascular cuffs. In CER, myelin lesions are usually bilaterally symmetrical and most severe in the occipital lobes, but they become more asymmetrical as they advance toward the frontal lobes⁵³. Exceptions to this rule even within the same family are well documented⁸.

Defects in CNS myelin are common in AMN. In contrast to CER, the lesions most commonly consist of foci of myelin pallor and loss with minimal reactive astrocytosis and lymphocytic infiltrates⁴⁸. Sparse collections of macrophages, often striated and containing angulate lysosomes, are the most conspicuous cell reaction. SCHAUMBURG et al.⁵³ proposed that the major site of damage in AMN, rather than being the central or peripheral myelin, is more likely to be the axons of long spinal tracts. While some degree of myelinolysis and dysmyelination should occur concurrently with the axonal insult, in AMN this appears to usually remain a minor pathologic element⁴⁶. However, in a minority of AMN patients, usually after many years of suffering from their myelopathy, some myelinolytic lesions progress to the inflammatory demyelination typical of CER.

In the destruction of the other cell types affected in X-ALD, i.e., adrenocortical cells, interstitial cells of Leydig, and Schwann cells, there are almost no inflammatory infiltrates. Free fatty acids are toxic to a variety of cells and their toxicity appears to increase with chain length. Thus, a cytotoxic pathogenesis has been pro-

posed for the adrenal, testicular and Schwann cell lesions in CER and AMN⁴⁸.

Cytokine Studies in X-ALD

The cerebral inflammatory reaction could be regulated at the level of inflammatory mediators (e.g. cytokines). Cytokines are antigen-nonspecific glycoproteins, synthesized and generally rapidly secreted in response to a stimulus. Cytokines are believed to act over a short range and to have very short half-lives. There are two distinct subtypes of CD4⁺ T helper (Th) cells, Th1 and Th2, that are identifiable because each produces a different set of cytokines. Th1 associated cytokines are interleukin (IL)-2, IL-12, interferon (IFN)- γ and lymphotoxin (TNF- β) and Th2 associated cytokines are IL-4, IL-5, IL-6 and IL-10. Th1 cells are the major players in delayed-type hypersensitivity and in pro-inflammatory responses by activating cytotoxic T cells and macrophages, while Th2 cells induce B cell growth and differentiation and induce antibody production. The anti-inflammatory cytokines produced by the Th2 cells can down-regulate Th1 cell function.

We investigated the nature of the inflammatory demyelinating reaction in X-ALD by comparing cytokine gene expression in cerebral inflammatory demyelinating lesions in X-ALD and MS in an attempt to define the inflammatory demyelinating process in X-ALD. Expression of mRNAs of the cytokines TNF- α , IL-1 β and IL-6 produced by macrophages (monokines), the lymphokines IFN- γ , IL-2, IL-4, IL-6 produced by T cells, and the TNF receptors [TNFRI (55 kDa) and TNFRII (75 kDa)] was measured in lesion samples taken *postmortem* from patients with CER and MS.

Our cytokine mRNA expression results for MS lesions confirmed immunocytochemical and *in situ* hybridization data in the literature showing that levels of the major inflammatory cytokines are elevated in MS brain lesions^{6, 15, 16, 54, 67, 68}. We showed that expression of monokines and lymphokines is generally increased in MS lesions when compared with CER lesions or controls. IL-4, associated with Th2 cells, is involved in down-regulation of the cellular immune response. IFN- γ , associated with Th1 cells, enhances the cellular immune response. We showed that CER lesion samples do not have IL-4 mRNA, while MS patients have IL-4 mRNA in both acute and gliotic lesion samples. Also, MS patients have a decrease in IFN- γ in gliotic samples compared with acute samples, while in CER there is an increase in IFN- γ in gliotic lesion samples compared with CER acute lesion samples.

In our studies, the level of expression of a number

of inflammatory cytokines in CER lesions is lower than that in MS lesions and more comparable with expression levels in control white matter samples. This suggests that transcriptional control of cytokine production in the inflammatory demyelinating reaction differs between CER and MS.

Tumor Necrosis Factor- α Studies in X-ALD

Several studies have implicated tumor necrosis factor- α (TNF- α), a cytokine, in the pathogenesis of a number of brain disorders, e.g. MS^{15, 54}, acquired immune deficiency syndrome (AIDS) encephalopathy^{34, 66}, bacterial meningitis⁴¹, cerebral malaria³¹ and X-ALD²⁹. TNF- α is an important mediator of inflammation and has been shown to be capable of selectively damaging oligodendrocytes and myelin sheaths *in vitro*⁵⁴. There is some controversy as to the significance of TNF- α in MS. Several groups have found elevated levels of TNF- α in cerebrospinal fluid and sera from MS patients^{7, 13, 28, 56, 63}. Therapies aimed at down-regulating TNF- α appear promising for the treatment of MS²⁰. However, other investigators found that the frequency of detectable cerebrospinal fluid and serum TNF- α was similar in MS patients and controls^{10, 44, 45}.

POWERS et al.⁴⁸ have shown that in acute inflammatory lesions from *postmortem* brain tissue of X-ALD patients with cerebral involvement, reactive astrocytes are most immunoreactive for TNF- α with macrophages staining less intensely. This is unusual, as in most pathologic conditions TNF- α production is associated primarily with macrophages. Astrocytes are most immunoreactive for TNF- α at the active demyelinating edge of the X-ALD lesion, with the inflammatory infiltrates (T cells and macrophages) located behind the active edge and in subsequent secondary tract degeneration. In MS, astrocytes are also most immunoreactive for TNF- α ^{15, 54} at the active demyelinating lesion edge, but in MS the inflammatory infiltrates are also located at the active lesion edge⁴⁸. These data suggest that TNF- α immunoreactive astrocytes, at the lesion edge, may play a primary role in the inflammatory demyelinating process in CER. The prevalence of TNF- α positive astrocytes at the active demyelinating lesion edge in X-ALD, and even beyond in otherwise morphologically normal white matter, indicates some pathogenetic connection⁴⁸. The observed decrease in astrocytic immunoreactivity in late acute to chronic active X-ALD lesions provides additional circumstantial evidence for a role in the early phase of demyelination.

In our laboratory we investigated TNF- α as a candidate modifying gene in X-ALD. We reported an in-

crease in TNF- α bioactivity in serum from patients with severe CCER. However, neither allelic differences in TNF- α nor levels of soluble TNF receptors accounted for the bioactivity differences or phenotypic heterogeneity in X-ALD²⁹.

In the earlier studies of POWERS et al.⁴⁸ it was unknown whether TNF- α was synthesized in astrocytes, accumulated in astrocytes or bound to astrocytes in CER lesions. Surprisingly, we showed that the level of TNF- α mRNA in CER acute/late acute lesion samples is only slightly above the level of detection²⁹. However, using *in situ* hybridization, we demonstrated that TNF- α mRNA is localized within astrocytes and macrophages in CER patients²⁹. Immunocytochemical localization of TNF- α in astrocytes has also been reported in MS by SELMAJ et al.⁵⁴ and in AIDS by TYOR et al.⁶⁴, although it is primarily localized to macrophages. Failure to detect TNF- α mRNA in total RNA from lesion cross sections together with the clear demonstration of TNF- α protein⁴⁸ and TNF- α mRNA²⁹ in astrocytes at the active demyelinating edge of inflammatory lesions implies that TNF- α expression is limited to a few reactive cells that might be important in the initiation of demyelination in the CER form of X-ALD. Failure to find allelic differences suggests that TNF- α does not account for the cerebral inflammatory response in X-ALD. Production of TNF- α by astrocytes may be followed by activation of macrophages and involvement of T cells in CER and progression of localized demyelination.

Human Leukocyte Antigen (HLA) Studies in X-ALD

Associations between HLA class I and class II alleles and various diseases have been reported by many investigators, e.g. in MS¹¹, systemic lupus erythematosus¹⁴ and rheumatoid arthritis⁴². It is possible that in some X-ALD individuals an altered metabolite of ALDP (an endogenously synthesized antigen) might be presented by specific HLA class I molecules to CD8⁺ cytotoxic T cells and thereby initiate an inflammatory reaction. Alternatively, if increased VLCFA in X-ALD brain tissue resulted in instability of myelin and subsequent myelinolysis, in some individuals, some of these myelin breakdown protein products (as exogenous antigens) might be presented by specific HLA class II molecules to CD4⁺ T helper cells, thus initiating the inflammatory reaction. We studied HLA association within the major X-ALD phenotypes (CCER and AMN) by typing HLA class I and class II specificities on lymphoblastoid cells from X-ALD patients and controls to determine whether or not there is an association

between HLA haplotype and X-ALD phenotype and compared our results to published findings in MS, the prototypical inflammatory demyelinating disorder³⁰. However, when we compared HLA specificities in CCER and AMN we found no statistically significant associations with phenotype. Thus, HLA is not likely to be a primary determinant of phenotypic heterogeneity in this disease³⁰.

Biochemical Abnormalities in CER Brain

It has been proposed that VLCFA excess in brain tissue from patients with the cerebral form of X-ALD acts as the biochemical trigger for the inflammatory response, but the exact nature of this trigger has yet to be defined. One of the difficulties in these studies has been the lack of availability of non-affected AMN brain tissue for comparison with non-affected CER brain tissue. Whereas a mild to moderate excess of VLCFA is present in all brain lipids in the CER form of X-ALD, the greatest excess occurs in the cholesterol ester, phosphatidylcholine (PC), ganglioside and proteolipid protein (PLP) fractions. Several authors have reported that there is an increased amount of cholesterol ester with an excess of VLCFA (C25:0, C26:0) in demyelinated regions of X-ALD brain^{5, 17, 18, 23, 32, 35, 49, 50, 51, 61}. IGARASHI et al.¹⁸, REINECKE et al.⁵² and THEDA et al.⁶² found a correlation between VLCFA level and degree of histologic abnormality in CCER brain and concluded that the cholesterol ester abnormality is a secondary phenomenon related to breakdown of myelin and subsequent release of free fatty acids, which become esterified with cholesterol and may serve a protective function. It is thus unlikely that cholesterol esterified with VLCFA acts as a trigger for the inflammatory response. Furthermore, OGINO and SUZUKI⁴³ showed that the degradation of cholesterol esters, including those that contain VLCFA, was unimpaired in X-ALD.

THEDA et al.⁶² reported an excess in the C26:0 content of the PC fraction from apparently intact regions of brain tissue from one CCER patient. PC is a major constituent of white matter and myelin lipid in normal brain and is probably positioned on the external surface of the myelin sheath⁶². Location on the external surface of the myelin sheath would make a modified PC more accessible for recognition by the immune system. Neurologic disorders associated with anti-phospholipid antibodies have been described, but with clinical and neuropathological findings that differ from those in CCER.

VLCFA account for 28 to 50% of the fatty acids in X-ALD brain gangliosides in moderately affected white matter, compared with 2.5% in normal brain^{17, 23}. C24:0

and C24:1 (and not C25:0 and C26:0) represent the predominant VLCFA in X-ALD gangliosides. LADISH et al.²⁶ showed that the length of the fatty acyl chain of the ceramide moiety dramatically influences ganglioside immunosuppressive activity. Gangliosides with shorter fatty acyl chains (C16:0, C18:0) are far more potent in inhibiting the *in vitro* lymphoproliferative response to a soluble antigen than are those containing longer fatty acyl chains (C22:0, C24:0, C24:1). Furthermore, unlike phospholipid components of membranes, gangliosides potentially can partition freely between membranes and the aqueous environment and thus passively migrate between cells. This type of ganglioside translocation could act as a physiological modulator of immune responses. Because some gangliosides, e.g. GM4, and PLP are unique to the CNS, i.e., not found in the PNS, and because the inflammatory response in the CER form of X-ALD is in the CNS only, gangliosides are potential triggers of the inflammatory response in the CER form of X-ALD.

In myelin, from both normally appearing and affected white matter from CCER brain, the proportion of saturated and mono-unsaturated VLCFA bound to PLP was increased at the expense of oleic acid (C18:0)³. PLP has recently emerged as a significant antigen in the prototypic demyelinating disease, MS, and as a potent encephalitogen in experimental models⁵⁹. Inter-patient heterogeneity in PLP may contribute to phenotypic variation in X-ALD. One possible mechanism for this is alteration of the protein's antigenicity.

Conclusion

In summary, a number of candidate modifying genes in X-ALD (cytokines and HLAs) have been investigated, but none appear to account for the cerebral inflammatory demyelinating reaction and/or phenotypic heterogeneity observed. The pathogenetic mechanism of the cerebral inflammatory demyelinating reaction in X-ALD and the role of the protein product of the X-ALD gene at Xq28 in destructive demyelination are as yet unknown. The X-ALD protein, ALDP, is a peroxisomal membrane protein and a member of the ATP-binding cassette (ABC) transporter superfamily, ALDP is closely related to at least three other peroxisomal membrane proteins, PMP70¹⁹, the X-ALD related protein (ALDR)²⁷ and the PMP70 related protein (P70R)⁵⁵. ALDP forms homodimers and heterodimers with these other peroxisomal ABC proteins⁵⁷. The genes for these three ALDP-related peroxisomal membrane ABC-proteins and for VLCS⁶⁰ are currently being investigated as potential modifier genes in X-ALD.

References

1. ANTOKU Y., KOIKE F., OHTSUKA Y., SAKAI T., TSUKAMOTO K., NAGARA H., IWASHITA H. and GOTO I. (1991): Adrenoleukodystrophy. A correlation between saturated very long chain fatty acids in mononuclear cells and phenotype. *Ann. Neurol.*, **30**, 101–103.
2. BERGER J., MOLZER B., FAE I. and BERNHEIMER H. (1994): X-linked adrenoleukodystrophy (ALD). A novel mutation of the ALD gene in 6 members of a family presenting with 5 different phenotypes. *Biochem. Biophys. Res. Commun.*, **205**, 1638–1643.
3. BIZZOZERO O. A., ZUNIGA G. and LEES M. B. (1991): Fatty acid composition of human myelin proteolipid protein in peroxisomal disorders. *J. Neurochem.*, **56**, 872–878.
4. BOLES D. J., CRAFT D. A., PADGETT D. A., LORIA R. M. and RIZZO W. B. (1991): Clinical variation in X-linked adrenoleukodystrophy – fatty acid and lipid metabolism in cultured fibroblasts. *Biochem. Med. Metab. Biol.*, **45**, 74–91.
5. BROWN III F. R., CHEN W. W., KIRSCHNER D. A., FRAYER K. L., POWERS J. M., MOSER A. B. and MOSER H. W. (1983): Myelin membrane from adrenoleukodystrophy brain white matter – isolation and physical/chemical properties. *J. Neurochem.*, **41**, 341–348.
6. CANNELLA B. and RAINE C. S. (1995): The adhesion molecule and cytokine profile of multiple sclerosis lesions. *Ann. Neurol.*, **37**, 424–435.
7. CHOFFLON M., JULLIARD C., JULLIARD P., GAUTHIER G. and GRAU G. E. (1992): Tumor necrosis factor- α production as a possible predictor of relapse in patients with multiple sclerosis. *Eur. Cytokine Netw.*, **3**, 523–531.
8. ESIRI M. M., HYMAN N. M., HORTON W. L. and LINDENBAUM R. H. (1984): Adrenoleukodystrophy. Clinical, pathological and biochemical findings in two brothers with the onset of cerebral disease in adult life. *Neuropathol. Appl. Neurobiol.*, **10**, 429–445.
9. FEIGENBAUM V., LOMBARD-PLATET G., GUIDOUX S., SARDE C., MANDEL J. L. and AUBOURG P. (1996): Mutational and protein analysis of patients and heterozygous women with X-linked adrenoleukodystrophy. *Am. J. Hum. Genet.*, **58**, 1135–1144.
10. FRANCIOTTA D. M., GRIMALDI L. M. E., MARTINO G. V., PICCOLO G., BERGAMASCHI R., CITTERIO A. and MELZI D'ERIL G. V. (1989): TNF in serum and CSF of patients with multiple sclerosis. *Ann. Neurol.*, **26**, 787–789.
11. FREEDMAN B. I., SPRAY B. J., HEISE E. R., ESPELAND M. A. and CANZANELLO V. J. (1993): A race-controlled human leukocyte antigen frequency analysis in lupus nephritis. The South-Eastern Organ Procurement Foundation. *Am. J. Kidney Dis.*, **21**, 378–382.
12. GRIFFIN D. E., MOSER H. W., MENDOZA Q., MOENCH T., O'TOOLE S. and MOSER A. B. (1985): Identification of the inflammatory cells in the nervous system of patients with adrenoleukodystrophy. *Ann. Neurol.*, **18**, 660–664.
13. HAUSER S. L., BHAN A. L., GILLES F., KEMP M., KERR C. and WEINER H. L. (1986): Immunohistochemical analysis of the cellular infiltrate in multiple sclerosis lesions. *Ann. Neurol.*, **19**, 578–587.
14. HILLERT J. (1994): Human leukocyte antigen studies in multiple sclerosis. *Ann. Neurol.*, **36**, S15–S17.

15. HOFMAN F. M., HINTON D. R., JOHNSON K. and MERRILL J. E. (1989): Tumor necrosis factor identified in multiple sclerosis brain. *J. Exp. Med.*, **170**, 607–610.
16. HOFMAN F. M., VAN HANWEHR R. I., DINARELLO C. A., MIZEL S. B., HINTON D. and MERRILL J. E. (1986): Immunoregulatory molecules and IL-2 receptors identified in multiple sclerosis brain. *J. Immunol.*, **136**, 3239–3245.
17. IGARASHI M., BELCHIS D. and SUZUKI K. (1976): Brain gangliosides in adrenoleukodystrophy. *J. Neurochem.*, **27**, 327–328.
18. IGARASHI M., SCHAUMBURG H. H., POWERS J., KISHIMOTO Y., KOLODNY E. and SUZUKI K. (1976): Fatty acid abnormality in adrenoleukodystrophy. *J. Neurochem.*, **26**, 851–860.
19. KAMIJO K., TAKETANI S., YOKOTA S., OSUMI T. and HASHIMOTO T. (1990): The 70-kDa peroxisomal membrane protein is a membrane of the Mdr (P-glycoprotein)-related ATP-binding protein superfamily. *J. Biol. Chem.*, **265**, 4534–4540.
20. KARIN N., MITHELL D., BROCKE S. and STEINMAN L. (1994): Reversal of experimental autoimmune encephalomyelitis by a soluble variant of a myelin basic protein epitope: T cell receptor antagonism and reduction of interferon- γ and TNF- α production. *J. Exp. Med.*, **180**, 2227–2237.
21. KEMP S., LIGTENBERG J. L., VAN GEEL B. M., BARTH P. G., WOLTERMAN R. A., SCHOUTE F., SARDE C.-O., MANDEL J. L., VAN OOST B. A. and BOLHUIS P. A. (1994): Identification of a two base pair deletion in five unrelated families with adrenoleukodystrophy. A possible hot spot for mutations. *Biochem. Biophys. Res. Commun.*, **202**, 647–653.
22. KEMP S., MOOYER P. A. W., BOLHUIS P. A., VAN GEEL B. M., MANDEL J. L., BARTH P. G., AUBOURG P. and WANDERS R. J. A. (1996): ALDP expression in fibroblasts of patients with X-linked adrenoleukodystrophy. *J. Inherit. Metab. Dis.*, **19**, 667–674.
23. KISHIMOTO Y., MOSER H. W. and SUZUKI K. (1984): Neurochemistry of adrenoleukodystrophy. In LAJTHA A. E. (ed.): *The handbook of neurochemistry*. Plenum Press, New York, **10**, 125–151.
24. KOK F., NEUMANN S., SARDE C.-O., ZHENG S., WU K.-H., BERGIN J., WATKINS P. A., GOULD S., SACK G., MOSER H. W., MANDEL J. L. and SMITH K. D. (1995): Mutational analysis of patients with X-linked adrenoleukodystrophy. *Hum. Mutat.*, **6**, 104–115.
25. KUMAR A. J., KOEHLER W., KRUSE B., NAIDU S., BERGIN A., EDWIN D. and MOSER H. W. (1995): MR findings in adult-onset adrenoleukodystrophy. *Am. J. Neuroradiol.*, **16**, 1227–1237.
26. LADISH S., LI R. and OLSON E. (1994): Ceramide structure predicts tumor ganglioside immunosuppressive activity. *Proc. Natl. Acad. Sci. USA*, **91**, 1974–1978.
27. LOMBARD-PLATET G., SAVARY S., SARDE C.-O., MANDEL J. L. and CHIMINI G. (1996): A close relative of the adrenoleukodystrophy (ALD) gene codes for a peroxisomal protein with a specific expression pattern. *Proc. Natl. Acad. Sci. USA*, **93**, 1265–1269.
28. MAIMONE D., GREGOR S., ARNASON B. G. and REDER A. T. (1991): Cytokine levels in CSF and serum of patients with MS. *J. Neuroimmunol.*, **32**, 67–74.
29. MCGUINNESS M. C., GRIFFIN D. E., RAYMOND G. V., WASHINGTON C. A., MOSER H. W. and SMITH K. D. (1995): Tumor necrosis factor- α and X-linked adrenoleukodystrophy. *J. Neuroimmunol.*, **61**, 161–169.
30. MCGUINNESS M. C., POWERS J. M., BIAS W. B., SCHMECKPEPER B. J., SEGAL A. H., GOWDA V. C., WESSELINGH S. L., BERGER J., GRIFFIN D. E. and SMITH K. D. (1997): Human leukocyte antigens and cytokine expression in cerebral inflammatory demyelinating lesions of X-linked adrenoleukodystrophy and multiple sclerosis. *J. Neuroimmunol.*, **75**, 174–182.
31. MCGUIRE W., HILL A. V. S., ALLSOP C. E. M., GREENWOOD B. M. and KWJATKOWSKI D. (1994): Variation in the TNF- α promoter region associated with susceptibility to cerebral malaria. *Nature*, **371**, 508–511.
32. MENKES J. H. and CORBO L. M. (1977): Adrenoleukodystrophy. Accumulation of cholesterol esters with very long chain fatty acids. *Neurology*, **27**, 928–932.
33. MIGEON B. R., MOSER H. W., MOSER A. B., AXELMAN J., SILLENCE D. and NORUM R. A. (1981): Adrenoleukodystrophy. Evidence for X-linkage, inactivation and selection favoring the mutant allele in heterozygous cells. *Proc. Natl. Acad. Sci. USA*, **78**, 5066–5070.
34. MINTZ M., RAPAPORT R., OLESKE J. M., CONNOR E. M., KOENIGSBERGER M. R., DENNY T. and EPSTEIN L. G. (1989): Elevated serum levels of tumor necrosis factor are associated with progressive encephalopathy in children with acquired immunodeficiency syndrome. *Am. J. Dis. Child.*, **143**, 771–774.
35. MOLZER B., BERNHEIMER H., BUDKA H., PILZ P. and TOIFL K. (1981): Accumulation of very long chain fatty acids is common to 3 variants of adrenoleukodystrophy. *J. Neurol. Sci.*, **51**, 301–310.
36. MOSER H. W., MOSER A. B., SINGH I. and O'NEILL B. R. (1984): Adrenoleukodystrophy. Survey of 303 cases: biochemistry, diagnosis and therapy. *Ann. Neurol.*, **16**, 628–641.
37. MOSER H. W., MOSER A. B., SMITH K. D., BERGIN A., BOREL J., SHANKROFF J., STINE O. C., MERETTE C., OTT J., KRIVIT W. and SHAPIRO E. (1992): Adrenoleukodystrophy. Phenotypic variability: implications for therapy. *J. Inherit. Metab. Dis.*, **15**, 645–664.
38. MOSER H. W., SMITH K. D. and MOSER A. B. (1994): X-linked adrenoleukodystrophy. In SCRIVER C. R., BEAUDET A. L., SLY W. S. and VALLE D. E. (eds.): *The metabolic and molecular basis of inherited disease*. 7th ed. McGraw Hill, New York, 2325–2349.
39. MOSSER J., DOUAR A.-M., SARDE C.-O., KIOSCHIS P., FEIL R., MOSER H. W., POUSTKA A.-M., MANDEL J.-M. and AUBOURG P. (1993): Putative X-linked adrenoleukodystrophy gene shares unexpected homology with ABC transporters. *Nature*, **361**, 726–730.
40. MOSSER J., LUTZ Y., STOECKEL M. E., SARDE C.-O., KRETZ C., DOUAR A. M., LOPEZ J., AUBOURG P. and MANDEL J. L. (1994): The gene responsible for adrenoleukodystrophy encodes a peroxisomal membrane protein. *Hum. Mol. Genet.*, **3**, 265–271.
41. MUSTAFA M. M., RAMILO O., OLSEN K. D., FRANKLIN P. S., HANSEN E. J., BEUTLER B. and MCCracken G. H. JR. (1989): Tumor necrosis factor in mediating experimental haemophilus influenza type B meningitis. *J. Clin. Invest.*, **84**, 1253–1259.
42. NEPOM G. T. and NEPOM B. S. (1992): Prediction of susceptibility to rheumatoid arthritis by human leukocyte antigen genotyping. *Rheum. Dis. Clin. North Am.*, **18**, 785–792.
43. OGINO T. and SUZUKI K. (1981): Specificities of human and rat brain enzymes of cholesterol ester metabolism toward very long chain fatty acids. Implications for biochemical pathogenesis of adrenoleukodystrophy. *J. Neurochem.*, **36**, 776–779.

44. PARRELLA O., CARRIERI P. B., DE MERCATO R. and BUSCAINO G. A. (1993): Markers of activated T-lymphocytes and T-cell receptor $\gamma\delta^+$ in patients with MS. *Eur. Neurol.*, **33**, 152–155.
45. PETER G. B., BOCTOR F. N. and TOURTELLOTE W. W. (1991): Serum and CSF levels of IL-2, sIL-2r, TNF α , and IL-1 β in chronic progressive multiple sclerosis. Expected lack of clinical utility. *Neurology*, **41**, 121–123.
46. POWERS J. M. (1985): Adrenoleukodystrophy (Adreno-testiculo-leuko-myelo-neuropathic-complex). *Clin. Neuropathol.*, **4**, 181–199.
47. POWERS J. M. (1994): The pathology of peroxisomal disorders with pathogenetic considerations. *J. Neuropathol. Exp. Neurol.*, **54**, 710–719.
48. POWERS J. M., LIU Y., MOSER A. B. and MOSER H. W. (1992): The inflammatory myelinopathy of adrenoleukodystrophy. Cells, effector molecules, and pathogenetic implications. *J. Neuropathol. Exp. Neurol.*, **51**, 630–643.
49. RAMSEY R. B., BANIK N. L. and DAVISON A. N. (1977): Galactolipid fatty acid composition in adrenoleukodystrophy. *J. Neurol. Sci.*, **32**, 69–77.
50. RAMSEY R. B., BANIK N. L. and DAVISON A. N. (1979): Adrenoleukodystrophy brain cholesteryl esters and other neutral lipids. *J. Neurol. Sci.*, **40**, 189–196.
51. RAMSEY R. B., BANIK N. L., SCOTT T. and DAVISON A. N. (1976): Neurochemical findings in adrenoleukodystrophy. *J. Neurol. Sci.*, **29**, 277–294.
52. REINECKE C. J., KNOLL D. P., PRETORIUS P. J., STEYN H. S. and SIMPSON R. H. W. (1985): The correlation between biochemical and histopathological findings in adrenoleukodystrophy. *J. Neurol. Sci.*, **70**, 21–38.
53. SCHAUMBURG H. H., POWERS J. M., RAINE C. S., SUZUKI K. and RICHARDSON E. P. (1975): Adrenoleukodystrophy. A clinical and pathological study of 17 cases. *Arch. Neurol.*, **33**, 577–591.
54. SELMAJ K., RAINE C. S., CANNELLA B. and BROSAN C. F. (1991): Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. *J. Clin. Invest.*, **87**, 949–954.
55. SHANI N., STEEL G., DEAN M. and VALLE D. (1997): Identification of a fourth half ABC transporter in the human peroxisomal membrane. *Hum. Mol. Genet.*, **6**, 1925–1931.
56. SHARIEF M. K. and HENTGES R. (1991): Association between tumor necrosis factor- α and disease progression in patients with multiple sclerosis. *N. Engl. J. Med.*, **325**, 467–472.
57. SMITH K. D., KEMP S., BRAITERMAN L. T., LU J.-F., WEI H.-M., GERAGHTY M., STETTEN G., BERGIN J. S., PEVSNER J. and WATKINS P. A. (1999): X-linked adrenoleukodystrophy. Genes, mutations, and phenotypes. *Neurochem. Res.*, **24**, 511–525.
58. SMITH K. D., SACK G., BEATY T., BERGIN A., NAIDU S., MOSER A. B. and MOSER H. W. (1991): A genetic basis for the multiple phenotypes of X-linked adrenoleukodystrophy. *Am. J. Hum. Genet.*, **49**, 165.
59. SOBEL R. A., GREER J. M. and KUCHROO V. K. (1994): Auto-immune responses to myelin proteolipid protein. *Neurochem. Res.*, **19**, 915–921.
60. STEINBERG S. I., WANG S. J., BRAITERMAN L. T. and WATKINS P. A. (1998): Investigation of the functional relationship between human very long chain acyl-CoA synthetase and adrenoleukodystrophy protein. *Am. J. Hum. Genet.*, **63** (suppl.), A275.
61. TAKETOMI T., HARA A., KITAZAWA N., TAKADA K. and NAKAMURA H. (1987): An adult case of adrenoleukodystrophy with features of olivo-ponto-cerebellar atrophy. II. Lipid biochemical abnormalities. *Jpn. J. Exp. Med.*, **57**, 59–70.
62. THEDA C., MOSER A. B., POWERS J. M. and MOSER H. W. (1992): Phospholipids in X-linked adrenoleukodystrophy white matter – fatty acid abnormalities before the onset of demyelination. *J. Neurol. Sci.*, **110**, 195–204.
63. TSUKADA N., MIYAGI K., MATSUDA M. and YANAGISAWA N. (1991): Tumor necrosis factor and interleukin 1 in the CSF and sera of patients with multiple sclerosis. *J. Neurol. Sci.*, **102**, 230–234.
64. TYOR W. R., GLASS J. D., BAUMRIND N., MCARTHUR J. C., GRIFFIN J. W., BECKER P. S. and GRIFFIN D. E. (1993): Cytokine expression of macrophages in HIV-1-associated vacuolar myelopathy. *Neurology*, **43**, 1002–1009.
65. WATKINS P. A., GOULD S. J., SMITH M. A., BRAITERMAN L. T., WEI H. M., KOK F., MOSER A. B., MOSER H. W. and SMITH K. D. (1995): Altered expression of ALDP in X-linked adrenoleukodystrophy. *Am. J. Hum. Genet.*, **57**, 292–301.
66. WESSELINGH S. L., POWER C., GLASS J. D., TYOR W. R., MCARTHUR J. C., FARBER J. M., GRIFFIN J. W. and GRIFFIN D. E. (1993): Intracerebral cytokine messenger RNA expression in acquired immunodeficiency syndrome dementia. *Ann. Neurol.*, **33**, 576–582.
67. WOODROFFE M. N., BELLAMY A. S., FELDMAN M., DAVISON A. N. and CUZNER M. L. (1988): Immunocytochemical characterization of the immune reaction in the central nervous system in multiple sclerosis. *J. Neurol. Sci.*, **84**, 257–264.
68. WOODRUFFE M. N. and CUZNER M. L. (1993): Cytokine mRNA expression in inflammatory (MS) lesions: detection by non-radioactive in situ hybridization. *Cytokine*, **5**, 583–588.

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