



Review

Role of Immune-Derived Diffusible Mediators in AIDS-Associated Neurological Disorders

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Abstract. Neurologic abnormalities are common in HIV-1 infected patients and often represent the dominant clinical manifestation of pediatric AIDS. Although the neurological dysfunction has been directly related to CNS invasion by HIV-1, the pathogenesis of neurologic disorders remains unclear. This review will first discuss the spectrum of potential interactions between HIV-1 and neural (neuronal and glial) cells, in the face of experimental data. Next, we will focus on the role of immune-derived cytokines and other soluble compounds which have been proposed to act as neurotoxic mediators and appear to play a role in the pathogenesis of AIDS-associated neurodegeneration.

Key words: AIDS patients; neurological disorders; neural cells.

Introduction

After the initial description of the acquired immune deficiency syndrome (AIDS), a slowly progressive, dementing illness was recognized in association with HIV-1 infection. Epidemiological studies have estimated that approximately one-third of patients with AIDS-related complex or Kaposi's sarcoma-defined AIDS have evidence of HIV-1 encephalopathy and another one-fourth have subclinical involvement^{36, 44}. HIV-1 encephalopathy is often characterized by an insidious onset of a disturbance in intellect. Fatigue and malaise, headaches and increasing social isolation are commonly observed. Job performance declines and self-care may become problematic^{1, 44}. HIV-1 encephalopathy generally occurs in the context of advanced immune suppression and coexistent systemic disease though it may represent the presenting and even sole

manifestation of the disease, particularly in children^{4, 23, 24} where it is accompanied by developmental delay or the loss of motor and intellectual milestones¹⁰. The gross pathology of HIV-1 encephalopathy is characterized by brain atrophy with sulcal widening and ventricular dilatation. Histologically, the presence of microglial nodules with multinucleated giant cells, diffuse astrogliosis, perivascular mononuclear inflammation, rarefaction of the white matter with microvacuolation, and neuronal loss⁹ characterize the lesions. Although the neurological dysfunction has been directly related to CNS invasion by HIV-1^{16, 35, 37, 40, 46, 50, 59}, the pathogenesis of neurologic disorders remains unclear.

Schematically, two mechanisms, not mutually exclusive, can be hypothesized which involve either direct neurocytopathic effects (depending upon HIV-1 infection of neuronal and/or glial cells) or indirect effects (mediated by HIV-1 infected and/or functionally acti-

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vated accessory and/or inflammatory cells). At present, the potential involvement of infectious agents other than HIV-1 acting as predisposing or inducing co-factors in the pathogenesis of AIDS-associated neurodegeneration remains to be clarified.

HIV-1 Neurotropic Properties: Direct Interactions with Specific Cell Types in the Brain

HIV-1 strain-specific properties may play a role in AIDS-associated neurodegeneration¹. In fact, HIV-1 neurotropism may, at least in part, depend upon the ability of the virus to enter and replicate in specific target cells (Fig. 1). These events depend upon cellular

and molecular interactions that require the expression of specific surface receptors by metabolically and transcriptionally active cells. Recent studies indicated that, in addition to the CD4 molecule, different chemokine receptors such as CXCR4, CCR5 and CCR3 could mediate virus entry in target cells¹³⁻¹⁵ (Fig. 1). These molecules are expressed in a broad range of tissues and cell types, including the brain and T cells. In particular, CCR3 is expressed by microglia in primary brain cultures³², while CXCR4 can be expressed by neurons, possibly playing a part in neurodevelopment⁷². Thus, M-tropic HIV-1 strains that use CCR5 or CCR3 may preferentially infect microglial cells, whereas T-tropic strains that use CXCR4 are unable to enter these cells and may potentially interact with neuronal cells³³. The CCR3 ligand, eotaxin, anti-CCR3 antibodies as well as

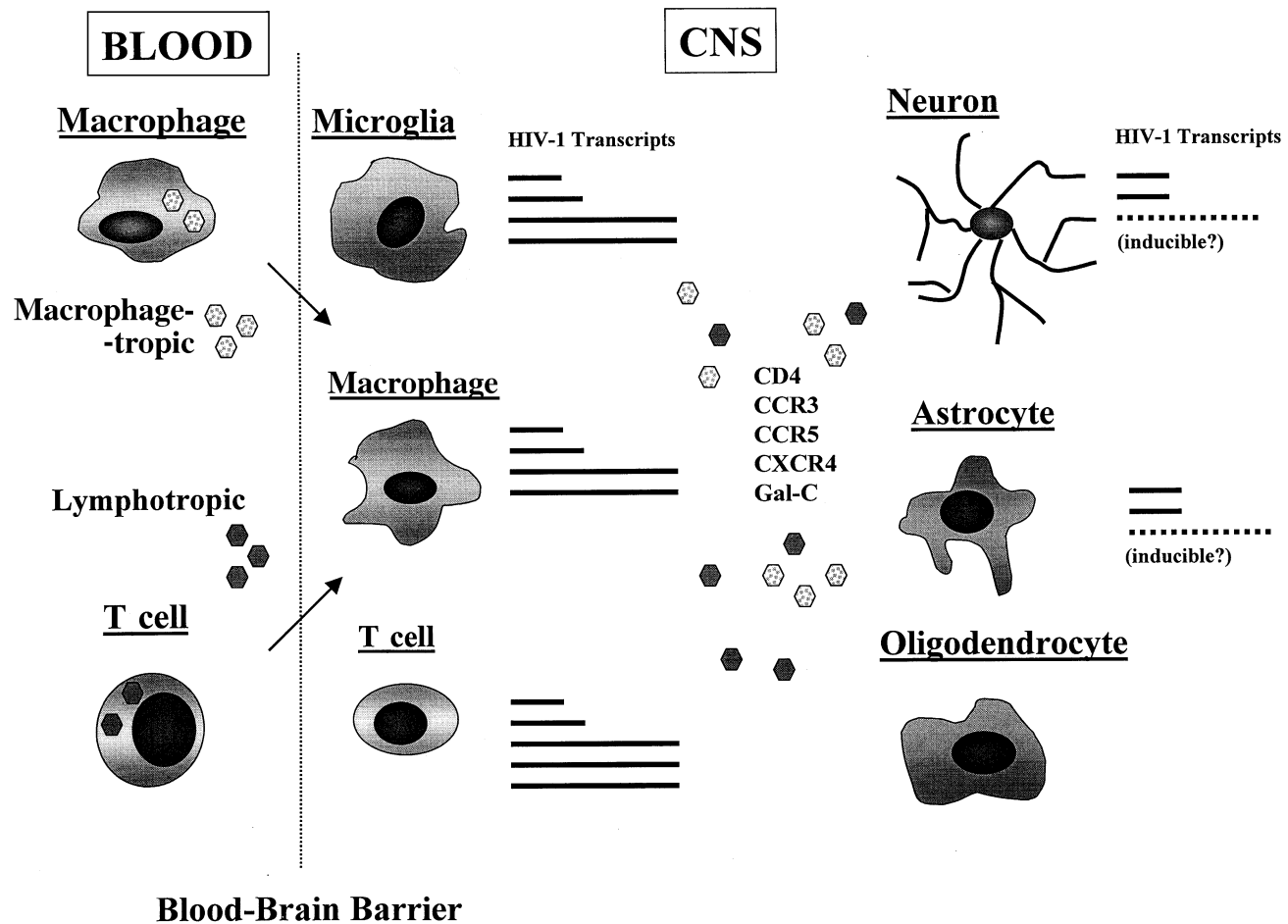


Fig. 1. Mechanisms controlling HIV-1 infection and replication in the brain. The distribution of molecules that can mediate HIV-1 binding and/or viral entry in target cells represents a first level of restriction of HIV-1 infection in the brain. It should be noted that in addition to chemokine receptors, additional molecules, such as a galactosyl-cerebroside, may play a role in mediating HIV-1 entry in neural cells. A second level of restriction is exerted at the intracellular level and corresponds to the degree of permissivity to viral gene expression and replication of specific cell types in the CNS. This is based, at least in part, upon the different utilization of regulatory elements within the viral promoter by distinct cell types, thus determining preferential environments for viral replication in individual neuronal and glial cells. Soluble signals can further modulate such permissivity and contribute in supporting virus latency or virus replication in the CNS⁶⁴ (see text for further explanation)

the CCR5 ligand, MIP-1 β , can inhibit HIV-1 infection of microglia, suggesting that these molecules may represent a potential target for new therapeutic strategies directed at reducing virus spreading in different tissues, including the brain.

HIV-1 infected neuronal and glial cells, however, have been rarely detected *in vivo* by conventional immunohistochemical techniques^{50, 44, 45}. Recently, the use of highly sensitive techniques such as the *in situ*, polymerase chain reaction (PCR), provided evidence for HIV-1 infection of these cells, in both adults and children, though accompanied by a low-level viral gene expression^{2, 46, 62}. *In vitro*, many different neuronal and glial cell lines as well as primary neurons and astrocytes appear susceptible to HIV-1 infection through a CD4-independent pathway of virus entry which appears to involve a galactosyl-cerebroside molecule^{17, 18, 32, 39, 45, 58, 65} (Fig. 1). Interestingly, the susceptibility to HIV-1 infection and the control of viral gene expression and replication in neural cells appear to depend upon the state of cellular differentiation being higher with more immature precursors than with differentiated cells^{17, 18, 22, 70, 71}. This may have important implications for pediatric HIV-1 infection and suggests that variations in the cellular microenvironment, either lineage- or differentiation-dependent, may contribute to influencing the ability of HIV-1 to infect and replicate in the nervous system.

The molecular features of such restricted HIV-1 infection of neurons and glial cells have been studied *in vitro* with cells transfected with the HIV-1 LTR, which contains the cis-acting regions that control virus expression in infected cells⁵², or with infectious HIV-1 proviral DNA, thus bypassing restriction posed by cell binding and entry. The predominant species of mRNA in these cells are the multiply spliced 2 kb species associated with regulatory gene expression (*tat*, *rev* and *nef*)^{63, 65}. Larger molecular weight RNA species associated with structural gene expression are virtually absent (Fig. 1). It should be noted that such regulatory gene products are generally released by target cells as biologically active extracellular soluble forms and may thus participate in HIV-1 neuropathogenesis by directly altering neuronal and glial viability and function or by contributing to inflammatory cell recruitment in the brain^{38, 41, 53}. In addition, recent reports indicate that regulatory elements of the viral LTR are differentially utilized according to the specific neuronal or glial cell types^{18, 22, 45, 58}. This could account for the spectrum of post-entry, strain-associated differences in the levels of virus replication which is generally observed with neural (neuronal and glial) cells of different origin^{18, 22, 45, 63}.

With these cell types HIV-1 replicates at very low, if not undetectable, levels (Fig. 1) and does not appear to induce evident cytopathic effects^{17, 18, 32, 62–65}. Taken together, these observations suggest that: 1) HIV-1 infection of neural cells may contribute to establishing a virus reservoir in the CNS whose extent and susceptibility to conventional antiretroviral therapy (including the more recent highly active regimens) are, at present, unknown, and 2) mechanism(s) other than direct virus-target interaction and, possibly, additional components of the brain microenvironment are required to induce the conspicuous pathology and functional damage observed in these patients.

Role of Persistent Immune Activation and Local Production of Diffusible Factors with Neurotoxic Properties

HIV-1 infection in the brain parenchyma is not abundant and does not seem to correlate with the clinical severity of the disease^{7, 30, 31}. Such a limited extent of viral burden suggests that the dementia in AIDS patients might be due to indirect effects, such as those induced by soluble factors produced by resident and blood-derived mononuclear cells (microglia, monocyte-macrophages, lymphocytes), which compelling evidence indicates as major targets and principal reservoirs of the virus in the brain^{2, 37, 50, 67–69}. Consistent with this hypothesis, recent studies indicated that the number of macrophages in the CNS of patients with AIDS is a better correlate to the dementia than viral burden^{7, 30, 31}. In addition to the production of viral structural and regulatory proteins (i.e. gp120, Tat), which, as previously mentioned, have been shown to possess neurotoxic properties *in vitro* and in animal models^{6, 38, 53, 61}, HIV-1 infected and/or functionally activated mononuclear cells can produce a number of inflammatory cytokines and bioactive substances which may both concur in regulating HIV-1 gene expression and replication in the CNS⁶⁴ and in altering neural cell function and survival^{29, 49} (Fig. 2). A recent set of studies determined that soluble mediators with neurotoxic properties are spontaneously produced by lymphomonocytes from HIV-1 infected individuals^{19, 20, 48}. Similar neurotoxic potentials are also expressed by normal PBMC or purified monocyte/macrophage (M/M) preparation upon functional activation. In fact, cell culture supernatants from both HIV-1 infected and functionally activated PBMC or M/M appear capable of altering the growth and viability of immature neurons as well as the survival of mitotically quiescent neuronal cells^{19, 20}. These effects

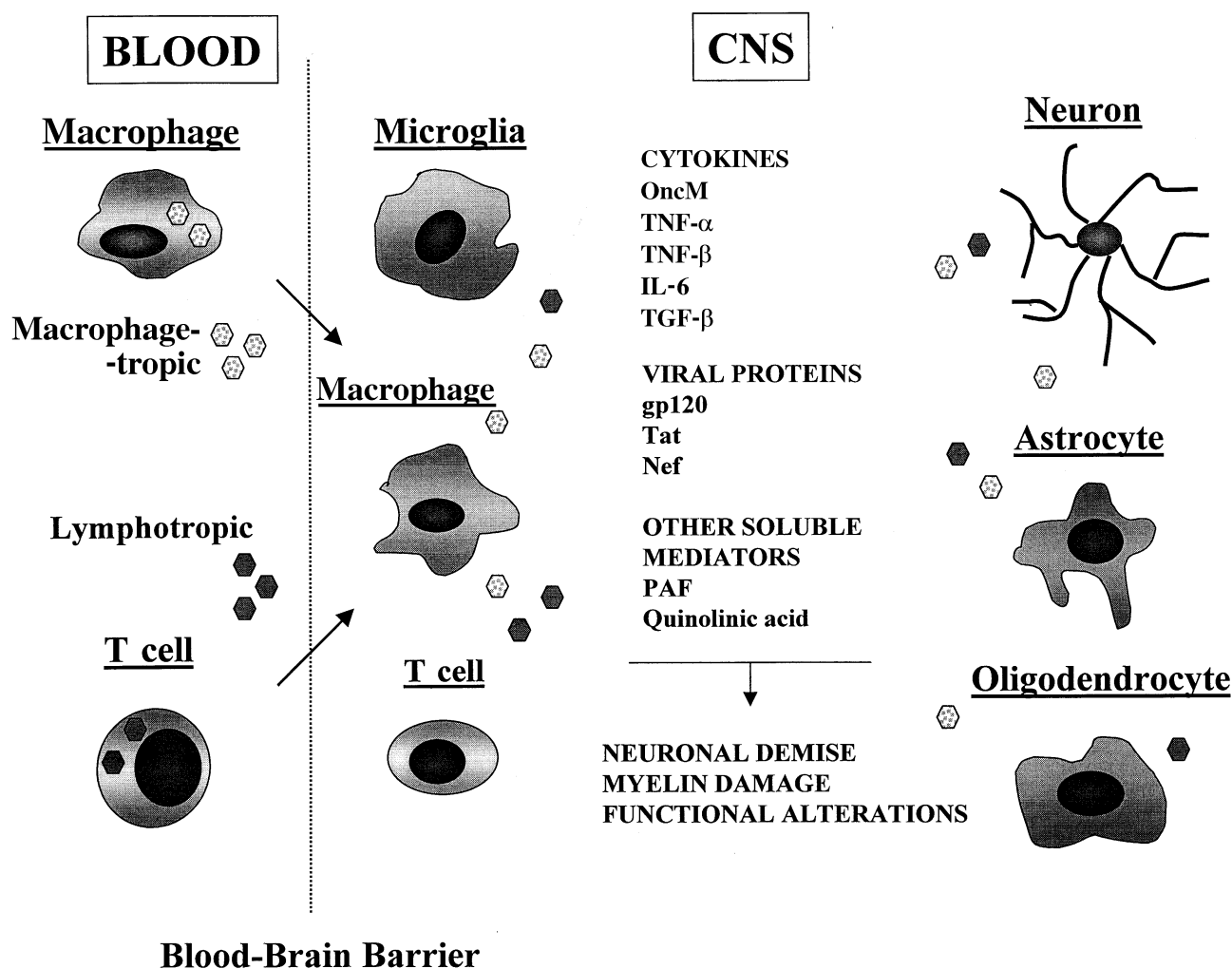


Fig. 2. Neuropathogenesis of HIV-1 infection. Potential contribution of viral proteins, inflammatory cytokines and neurotoxic metabolites in the pathophysiology of AIDS-associated neurodegeneration. Following CNS invasion by HIV-1 and immune cell activation and recruitment, multiple mechanisms, non-mutually exclusive, may participate in inducing neural damage and functional impairment (see text for further explanation)

are accompanied by both morphological and biochemical alterations of the cells consistent with the induction of apoptosis. Similar observations, obtained by distinct studies on different neural model systems^{28, 29, 48, 49}, suggest that both HIV-1 infection and functional activation can induce lymphomononuclear cells to express neurotoxic potentials which relay upon the production of bioactive substances that, in turn, induce neuronal growth perturbations and apoptotic cell death.

Several diffusible factors released by immune cells possess documented damaging potentials on neural tissues (Fig. 2). Among them, oncostatin M (oncM), a recently characterized proinflammatory mediator^{43, 51}, has been identified as one cytokine that directly causes a profound inhibition of neuronal proliferation and via-

bility. OncM belongs to a family of structurally and functionally related pleiotropic factors that utilize the gp130 or gp130-related receptor subunits on target cells^{43, 51}. These cytokines exhibit differential effects on a variety of cell types, including cells of neural, hematopoietic, lymphopoietic and vascular origin. Indeed, the same protein can act differentially on target cells, depending upon the responsive cell population²¹. Interestingly, expression of oncM in transgenic mice is detrimental to normal mouse development and death is associated with expression in neurons⁴², suggesting that oncM production in the brain may exert lethal effects on neuronal cells.

In the presence of oncM, the decrease in neuronal cell viability, documented by the altered membrane per-

meability to propidium iodide staining, is preceded and accompanied by extensive DNA cleavage which indicates that oncM induces apoptotic cell death. Moreover, antibodies neutralizing anti-oncM have been capable of preventing the growth inhibition induced by either HIV-1 infected or functionally activated lymphomonocytes as well as the effects of the recombinant protein²⁰.

Taken together, these data indicate that native oncM can play a part in HIV-1-associated neurodegeneration, though other factors are likely to be involved in the process, acting either in concert or independently from oncM. For instance, TGF- β_1 , but not TNF- α , IFN- γ , IL-6 or virus components such as gp120, can act in concert with oncM, enhancing the inhibitory effects of the cytokine on primary neurons and neuronal cell lines of different origins. In the presence of both cytokines, neuronal growth and survival alterations can be elicited at very low oncM concentrations, consistent with those spontaneously released by PBMC from HIV-1 infected subjects in their supernatants²⁰. Thus, an increased oncM production from HIV-1 infected or functionally-activated lymphocytes as well as resident or blood-derived mononuclear phagocytes can directly contribute to neuronal apoptosis in AIDS-associated neurodegeneration, which generally occurs in the absence of direct HIV-1 infection of these cells, and appears to depend upon the induction of diffusible factors^{47, 60}. These data also suggest that the total neurotoxic activity potentially exerted by HIV-1 infected or functionally activated lymphomononuclear cells depends upon multiple parameters that include the class of cytokine production, its concentration, potency and interaction(s) with other mediators.

In fact, several inflammatory mediators produced upon HIV-1 infection or functional activation of lymphomononuclear cells can play a part in the pathogenesis of HIV-1 induced neurodegeneration (Fig. 2). Altered levels of proinflammatory cytokines such as IL-1, IL-6, TNF- α and IFN- γ and the presence of neurotoxic metabolites such as quinolinic acid (a monocyte/microglial byproduct of tryptophan with neurotoxic properties) have been documented in sera and CSF of AIDS patients^{7, 25, 27, 28, 34, 48, 49}. Production of the same cytokines as well as TGF- β and arachidonic acid metabolites have also been documented in the brain parenchyma, in association with HIV-1 infected and/or activated mononuclear cells and astrocytes^{28, 46, 66}. Such soluble factors possess documented damaging potentials on neural tissues. In addition to mediating microglia and astrocytic activation, which in turn lead to increased proinflammatory cytokines production²¹, many of them can exert either a positive or a negative regulation of

oligodendrocytes proliferation and survival^{3, 54}. TNF- α has been shown to mediate myelin and oligodendrocytes damage both *in vivo* and *in vitro*^{55, 56} and to exert cytotoxic activity against rat oligodendrocytes, which results in cell death. TNF- β , which is genetically and functionally related to TNF- α , has also been shown to possess potent cytotoxicity against oligodendrocytes. The effect is much more potent than TNF- α and is mediated by apoptotic mechanisms⁵⁷. In addition to the previously mentioned effects of IL-1, TNF- α and TNF- β on oligodendrocyte survival and myelination, IL-6 has been shown to mediate potent neurotoxic effects when expressed in the brain of transgenic mice¹¹. Other soluble mediators such as the platelet-activating factor (PAF) and leukotriens/prostaglandins, which are produced upon microglia/macrophages and astrocytes functional interaction, have been reported to exert damaging effects on primary neural cells and neural cell lines^{27, 28}. Thus, in the event of lymphoid/monocytic cell infiltration into the CNS and the activation of resident microglia and astrocytes, the complex circuitry of molecular interactions mediated by cytokines appears altered both spatially and temporally and capable of mediating neurotoxic effects. In addition, the pleiotropic properties of such soluble mediators are responsible for multiple and apparently contrasting effects exerted upon different cell types in the brain. For example, TNF- α can either exert direct inhibitory or stimulatory effects depending upon the responsive neural cell type^{5, 21, 26}. In fact, TNF- α can even protect primary neurons of different origins from metabolic-excitotoxic insults¹². This is not surprising, since cytokines such as TNF- α , TGF- β , oncM and IL-6, key regulatory factors in the host response to injury or immunological challenge, can also act as instructive/permmissive signals during nervous system development or in events which involve functional or structural tissue remodeling²¹. In addition to direct interaction with responsive neuronal cell targets, the contribution of these cytokines to neuronal demise may involve multiple interactions with all the different non-neuronal cell types in the brain. This notion further emphasizes the concept that the inappropriate, chronic production of different inflammatory cytokines such as TNF- α , TGF- β , oncM and IL-6, which have partially overlapping effects and the potential to exert both direct and indirect CNS injury, can play a major role in the pathogenesis of neurologic AIDS-associated disorders by acting through different mechanisms on different target cells and potentially cooperate by either additive or synergistic modality, thus amplifying their effects on responsive cell types.

Thus, AIDS-associated neurodegeneration appears a complex, multifactorial disease where the combined action of HIV-1 infection, production of viral proteins and soluble inflammatory mediators are implicated in the pathophysiology (Fig. 2).

Concluding Remarks

Several lines of evidence suggest that a cascade of events triggered by HIV-1 infection and involving a chronic dysregulation of cytokine expression are implicated in the setting of neuronal demise in both the immature and adult brain. These findings contribute to identifying novel pathways of immunologically-mediated neuronal injury and may provide a model for the interpretation of neurological disorders depending upon CNS inflammatory conditions from varied insults. This evidence should be taken into account in the development of therapeutic intervention, which should be directed not only toward reducing the viral load in the brain, but also in controlling the deleterious effects of cytokines and protecting neural cells from toxic metabolites.

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References

1. ATWOOD W. J., BERGER J., KADERMAN R., TORNATORE C. S., and MAJOR E. O. (1993): Human immunodeficiency virus type-1 infection of the brain. *Clin. Microbiol. Rev.*, **6**, 339–366.
2. BAGASRA O., LAVI E., BOBROSKI L., KHALILI K., PESTANER J. P., TAWADROS R. and POMERANTZ R. J. (1996): Cellular reservoirs of HIV-1 in the central nervous system of infected individuals: identification by the combination of *in situ* polymerase chain reaction and immunohistochemistry. *AIDS*, **10**, 573–585.
3. BARRES, B. A., HART I. K., COLES H. S. R., BURNE J. F., VOYVODIC J. T., RICHARDSON W. D. and RAFF M. C. (1992): Cell death and control cell survival in the oligodendrocyte lineage. *Cell*, **70**, 31–46.
4. BELMAN A. L., ULTMAN M., KURTZBERG D., and CONE-WES-SON B. (1985): Neurological complications in children with AIDS. *Ann. Neurol.*, **18**, 560–566.
5. BOGDAN I., LEIB S. L., BERGERON M., CHOW L. and TAUBER M. G. (1997): Tumor necrosis factor- α contributes to apoptosis in hippocampal neurons during experimental group B streptococcal meningitis. *J. Infect. Dis.*, **176**, 693–701.
6. BRENNEMAN D. E., MCCUNE S. K., MERVIS R. F. and HILL J. M. (1994): Gp 120 as an etiologic agent for NeuroAIDS: neurotoxicity and model systems. *Adv. Neuroimmunol.*, **4**, 157–165.
7. BREW B., ROSENBLUM M., CRONIN K. and PRICE R. W. (1995): AIDS dementia complex and HIV-1 brain infection: clinical-virological correlations. *Ann. Neurol.*, **38**, 755–762.
8. BROUWERS P., HEYES M. P., MOSS H. A., WOLTERS P. L., POPLACK D. G., MARKEY S. P. and PIZZO P. A. (1993): Quinolinic acid in the cerebrospinal fluid of children with symptomatic human immunodeficiency virus type 1 disease: relationships to clinical status and therapeutic response. *J. Infect. Dis.*, **168**, 1380–1386.
9. BUDKA H. (1989): Human immunodeficiency virus (HIV)-induced disease of the central nervous system: pathology and implications for pathogenesis. *Acta Neuropathol.*, **77**, 225–236.
10. CALVELLI T. A. and RUBINSTEIN A. (1990): Pediatric HIV infection: a review. *Immunodef. Rev.*, **2**, 83–127.
11. CAMPBELL I. L., ABRAHAM C. R., MASLIAH E., KEMPER P., INGLIS J. D., OLDSTONE M. B. A. and MUCKE L. (1993): Neurologic disease induced in transgenic mice by the cerebral overexpression of interleukin-6. *Proc. Natl. Acad. Sci. USA*, **90**, 10061–10065.
12. CHENG B., CHRISTAKOS S. and MATTSON M. P. (1994): Tumor necrosis factor protects neurons against metabolic-excitotoxic insults and promotes maintenance of calcium homeostasis. *Neuron*, **12**, 139–147.
13. CHOE H., FARZAN M., SUN Y., SULLIVAN N., ROLLINS B., PONATH P. D., WU L., MACKAY C. R., LAROSA G., NEWMAN W., GERARD N., GERARD C. and SODROSKI J. (1996): The β -chemokine receptors CCR3 and CCR5 facilitate infection by primary HIV-1 isolates. *Cell*, **85**, 1135–1148.
14. DENG H., LIU R., ELLMEIER W., CHOE S., UNUTMATZ D., BURKHART M., DI MARZIO P., MARMON S., SUTTON R. E., HILL C. M., DAVIS C. B., PEIPER S. C., SCHALL T. J., LITTMAN D. R. and LANDAU N. R. (1996): Identification of a major co-receptor for primary isolates of HIV-1. *Nature*, **381**, 661–666.
15. DORANZ B. J., RUCKER J., YI Y., SMYTH R. J., SAMSON M., PEIPER S. C., PARMENTIER M., COLLMAN R. G. and DOMS R. W. (1996): A dual-tropic primary isolate that uses fusin and the β -chemokine receptors CKR-5, CKR-3, and CKR-2 β as fusion co-factors. *Cell*, **85**, 1149–1158.
16. ELBOTT D. J., PERESS N., BURGER H., LA NEVE D., ORENSTEIN J., GENDELMAN H. E., SEIDMAN R. and WEISER B. (1989): Human immunodeficiency virus type 1 in spinal cords of acquired immunodeficiency syndrome patients with myelopathy: expression and replication in macrophages. *Proc. Natl. Acad. Sci. USA*, **86**, 3337–3341.
17. ENSOLI F., CAFARO A., FIORELLI V., VANNELLI B., ENSOLI B. and THIELE C. J. (1995): HIV-1 infection of primary human neuroblasts. *Virology*, **210**, 221–225.
18. ENSOLI F., ENSOLI B. and THIELE C. J. (1994): HIV-1 gene expression and replication in neuronal and glial cell lines with immature phenotype: effects of nerve growth factor. *Virology*, **200**, 668–676.
19. ENSOLI F., FIORELLI V., DECRISTOFARO R., SANTINI-MURATORI D., NOVI A., LUZI G. and AIUTI F. (1998): Inflammatory cytokines and HIV-1 associated neurodegeneration: role of oncostatin-M production by HIV-1 infected or functionally activated mononuclear cells. *Proceedings of the 12th International Conference on AIDS*, Monduzzi Editore, Bologna, **5**, 71–77.
20. ENSOLI F., FIORELLI V., DECRISTOFARO R., SANTINI-MURATORI D., NOVI A., VANNELLI B., THIELE C. J., LUZI G. and AIUTI F. (1999): Inflammatory cytokines and HIV-1-associated neurodegeneration: oncostatin-M produced by mononuclear cells from

- HIV-1-infected individuals induces apoptosis of primary neurons. *J. Immunol.*, **162**, 6268–6277.
21. ENSOLI F., FIORELLI V., SANTINI-MURATORI D., VINCENZI L., TOPINO S., NOVI A., LUZI G. and SIRIANNI M. C. (1999): Immune-derived cytokines in the nervous system: epigenetic instructive signal or neuropathogenic mediators? *Crit. Rev. Immunol.*, **19**, 97–116.
 22. ENSOLI F., WANG H., FIORELLI V., ZEICHNER S. L., LUZI G. and THIELE C. J. (1997): HIV-1 infection and the developing nervous system: lineage-specific regulation of viral gene expression and replication in distinct neuronal precursors. *J. Neurovirol.*, **3**, 290–298.
 23. EPSTEIN L. G., SHARER L. R., JOSHI V. V., FOJAS M. M., KOENIGSBERGER M. R. and OLESKE J. M. (1985): Progressive encephalopathy in children with acquired immunodeficiency syndrome. *Ann. Neurol.*, **17**, 488–496.
 24. EPSTEIN L. G., SHARER L. R., OLESKE J. M., CONNOR E. M., GOUDSMIT J., BAGDON L., ROBERT-GUROFF M. and KOENIGSBERGER M. R. (1986): Neurological manifestations of HIV infection in children with AIDS. *Pediatrics*, **78**, 678–687.
 25. FINCH C. E., LAPING N. J., MORGAN T. E., NYCHOLS N. R. and PASINETTI J. N. (1993): TGF- β_1 is an organizer of responses to neurodegeneration. *J. Cell. Biochem.*, **53**, 314–322.
 26. FINE S. M., ANGEL R. A., PERRY S. W., EPSTEIN L. G., ROTHSTEIN J. D., DEWHURST S. and GELBARD H. A. (1996): Tumor necrosis factor alpha inhibits glutamate uptake by primary human astrocytes: implications for pathogenesis of HIV-1 dementia. *J. Biol. Chem.*, **271**, 15303–15306.
 27. GELBARD H. A., NOTTET H. S. M. L., SWINDELLS S., JETT M., DZENKO K. A., GENIS P., WHITE R., WANG H. A., CHOI Y. B., ZHANG D., LIPTON S. A., TOURTELLOTT W. W., EPSTEIN L. G. and GENDELMAN H. E. (1994): Platelet-activating factor: a candidate human immunodeficiency virus type 1-induced neurotoxin. *J. Virol.*, **68**, 4628–4635.
 28. GENIS P., JETT M., BERNTON E. W., BOYLE T., GELBARD H. A., DZENKO K. A., KENE R. W., RESNICK L., MIZRACHI Y., VOLSKY D. J., EPSTEIN L. G. and GENDELMAN H. E. (1992): Cytokines and arachidonic metabolites produced during human immunodeficiency virus (HIV)-infected macrophage-astroglia interactions: implications for the neuropathogenesis of HIV disease. *J. Exp. Med.*, **176**, 1703–1718.
 29. GIULIAN D., VACA K. and NOONAN C. A. (1990): Secretion of neurotoxins by mononuclear phagocytes infected with HIV-1. *Science*, **250**, 1593–1596.
 30. GLASS J. D., FEDOR H., WESSELINGH S. L. and MCARTHUR J. C. (1995): Immunocytochemical quantification of human immunodeficiency virus in the brain: correlation with dementia. *Ann. Neurol.*, **38**, 755–762.
 31. GLASS J. D., WESSELINGH S. L., SELNES O. A. and MCARTHUR J. C. (1993): Clinical-neuropathological correlation in HIV-associated dementia. *Neurology*, **43**, 2230–2237.
 32. HAROUSE J. M., KUNSCH C., HARTLE H. T., LAUGHLIN M. A., HOXIE J. A., WIGDAHL B. and GONZALEZ-SCARANO F. (1989): CD4-independent infection of human neural cells by human immunodeficiency virus type 1. *J. Virol.*, **63**, 2527–2533.
 33. HE J., CHEN Y., FARZAN M., CHOE H., OHAGEN A., GARTNER S., BUSCIGLIO J., YANG X., HOFMANN W., NEWMAN W., MACKAY C. R., SODROSKY J. and GABUZDA D. (1997): CCR3 and CCR5 are co-receptors for HIV-1 infection of microglia. *Nature*, **385**, 645–649.
 34. HEYES M. P., BREW B. J. and MARTIN A. (1991): Quinolinic acid in cerebrospinal fluid and serum in HIV-1 infection: relationship to clinical and neurologic status. *Ann. Neurol.*, **29**, 202–209.
 35. HO D. D., ROTA T. R., SCHOOLEY R. T., KAPLAN J. C., ALLAN J. D., GROOPMAN J. E., RESNICK L., FELSENSTEIN D., ANDREWS C. A. and HIRSCH M. S. (1985): Isolation of HTLV-III from cerebrospinal fluid and neural tissues of patients with neurologic syndrome-related to the acquired immunodeficiency syndrome. *N. Engl. J. Med.*, **313**, 1493–1497.
 36. JANSSEN R. S., CORNBATH D. R., EPSTEIN L. G., MCARTHUR J. and PRICE R. W. (1989): Human immunodeficiency virus (HIV) infection and the nervous system: report from the American Academy of Neurology, AIDS task force. *Neurology*, **39**, 119–122.
 37. KOENIG S., GENDELMAN H. E., ORENSTEIN J. M., DAL CANTO M. C., PEZESHKPOUR G. H., YUNGBLUTH M., JANOTTA F., AKSAMIT A., MARTIN M. A. and FAUCI A. S. (1986): Detection of AIDS virus in macrophages in brain tissue from AIDS patients with encephalopathy. *Science*, **233**, 1089–1093.
 38. KOLSON D. L., BUCHHALTER J., COLLMAN R., HELLMIG B., FARRELL C. F., DEBOUCK C. and GONZALEZ-SCARANO F. (1993): HIV-1 Tat alters normal organization of neurons and astrocytes in primary rodent brain cell cultures: RGD sequence dependence. *AIDS Res. Hum. Retroviruses*, **9**, 677–685.
 39. LI X. L., MOUDGIL T., VINTERS H. V. and HO D. D. (1990): CD4-independent, productive infection of a neuronal cell line by human immunodeficiency virus type 1. *J. Virol.*, **64**, 1383–1387.
 40. LYMAN W. D., KRESS Y., KURE K., RASHBAUM W. K., RUBINSTEIN A. and SOEIRO R. (1990): Detection of HIV in fetal central nervous system tissue. *AIDS*, **4**, 917–920.
 41. MALE D., PRICE G., HUGHES C. and LANTOS P. (1990): Lymphocyte migration into brain modelled *in vitro*: control by lymphocyte activation, cytokines and antigen. *Cell. Immunol.*, **127**, 1–11.
 42. MALIK N., HAUGEN H. S., MODRELL B., SHOYAB M. and CLEGG C. H. (1995): Developmental abnormalities in mice transgenic for bovine oncostatin M. *Mol. Cell. Biol.*, **15**, 2349–2358.
 43. MALIK N., KALLESTED J. C., GUNDERSON N. L., AUSTIN S. D., NEUBAUER M. G., OCHS V., MARQUARDT H., ZARLING J. M., SHOYAB M., WEI C. M., LINSLEY P. S. and ROSE T. M. (1989): Molecular cloning, sequence analysis, and functional expression of a novel growth regulator, Oncostatin M. *Mol. Cell. Biol.*, **9**, 2847–2853.
 44. MCARTHUR J. C., HOOVER D. R., BACELLAR H., MILLER E. N., COHEN B. A., BECKER J. T., GRAHAM N. M. H., MCARTHUR J. H., SELNES O. A., JACOBSON L. P., VISSCHER B. R., CONCHA M. and SAAH A. (1993): Dementia in AIDS patients: incidence and risk factor. *Neurology*, **43**, 2245–2252.
 45. MCCARTHY M., HE J. and WOOD C. (1998): HIV-1 strain-associated variability in infection of primary neuroglia. *J. NeuroVirol.*, **4**, 80–89.
 46. NUOVO G. J., GALLERY F., MACCONNELL P. and BRAUN A. (1994): *In situ* detection of polymerase chain reaction-amplified HIV-1 nucleic acids and tumor necrosis factor-alpha RNA in the central nervous system. *Am. J. Pathol.*, **144**, 659–666.
 47. PETITO C. K. and ROBERTS B. (1995): Evidence of apoptotic cell death in HIV encephalitis. *Am. J. Pathol.*, **146**, 1121–1130.
 48. PULLIAM L., CLARKE J. A., MCGRATH M. S., MOORE D. and MCGUIRE D. (1996): Monokine products as predictors of AIDS dementia. *AIDS*, **10**, 1495–1500.

49. PULLIAM L., HERNDIER B. G., TANG N. M. and McGRATH M. S. (1991): Human immunodeficiency virus-infected macrophages produce soluble factors that cause histological and neurochemical alterations in cultured human brains. *J. Clin. Invest.* **87**, 503–512.
50. PUMAROLA-SUNE T., NAVIA B. A., CORDON-CARDO C., CHO E. S. and PRICE R. W. (1987): HIV antigen in the brains of patients with the AIDS dementia complex. *Ann. Neurol.* **21**, 490–496.
51. ROSE T. N. and BRUCE A. G. (1991): Oncostatin M is a member of a cytokine family that includes leukemia-inhibitory factor, granulocyte colony-stimulating factor, and interleukin 6. *Proc. Natl. Acad. Sci. USA*, **88**, 8641–8645.
52. ROSEN C. A., SODROSKY J. G. and HASELTINE W. A. (1985): Location of the cis-acting regulatory sequences in the human T cell lymphotropic virus type III (HTLV-III) long terminal repeat. *Cell*, **41**, 813–823.
53. SABATIER J. M., VIVES E., MABROUK K., BENJOUAD A., ROCHAT H., DUVAL A., HUE B. and BAHRAOUI E. (1991): Evidence for neurotoxic activity of Tat from human immunodeficiency virus type 1. *J. Virol.* **65**, 961–967.
54. SANETHO R., CHIAPPELLI F. and DE VELLIS J. (1987): Interleukin-2 inhibition of oligodendrocyte progenitor cell proliferation depends on expression of the TAC receptor. *J. Neurosci. Res.*, **18**, 147–154.
55. SELMAJ K. W. and RAINE C. S. (1988): Tumor necrosis factor mediates myelin and oligodendrocyte *in vitro*. *Ann. Neurol.* **23**, 339–346.
56. SELMAJ K. W., RAINE C. S., CANNELLA B. and BROSNAM C. F. (1991): Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. *J. Clin. Invest.* **87**, 949–954.
57. SELMAJ K. W., RAINE C. S., FAROOQ M., NORTON W. T. and BROSNAM C. F. (1991): Cytokine cytotoxicity against oligodendrocytes: apoptosis induced by lymphotoxin. *J. Immunol.* **147**, 1522–1529.
58. SHARPLESS N., GILBERT D., VANDERCAM B., ZHOU J. M., VERDIN E., RONNETT G., FRIEDMANN E. and DUBOIS-DALCQ M. (1992): The restricted nature of HIV-1 tropism for cultured neural cells. *Virology*, **191**, 813–825.
59. SHAW G. M., HARPER M. E., HAHN B. H., EPSTEIN L. G., GAJDUSEK D. C., PRICE R. W., NAVIA B. A., PETITO C. K., O'HARA C. J., GROOPMAN J. E., CHO E. S., OLESKE J. M., WONG-STAAAL F. and GALLO R. C. (1985): HTLV-III infection in brains of children and adults with AIDS encephalopathy. *Science*, **227**, 177–182.
60. SHI B., DE GIROLAMI U., HE J., WANG S., LORENZO A., BUSCIGLIO J. and GABUZDA D. (1996): Apoptosis induced by HIV-1 infection of the central nervous system. *J. Clin. Invest.* **98**, 1979–1990.
61. TOGGAS S. M., MASLIAH E., ROCKENSTEIN E. M. and MUCKE L. (1994): Central nervous system damage produced by expression of the HIV-1 coat protein gp 120 in transgenic mice. *Nature*, **367**, 188–193.
62. TORNATORE C., CHANDRA R., BERGER J. R. and MAJOR E. O. (1994): HIV-1 infection of subcortical astrocytes in the pediatric central nervous system. *Neurology*, **44**, 481–487.
63. TORNATORE C., MEYERS K., ATWOOD W., CONANT K. and MAJOR E. O. (1994): Temporal patterns of human immunodeficiency virus type 1 transcripts in human fetal astrocytes. *J. Virol.* **68**, 93–98.
64. TORNATORE C., NATH A., AMEMIYA K. and MAJOR E. O. (1991): Persistent human immunodeficiency virus type 1 infection in human fetal glial cells reactivated by T-cell factor(s) or by the cytokines tumor necrosis factor alpha and interleukin-1 beta. *J. Virol.* **65**, 6094–6102.
65. TRUCKENMILLER M. E., KULAGA H., COGGIANO M., WYATT R., SNYDER S. H. and SWEETNAM P. M. (1993): Human cortical neuronal cell line: a model for HIV-1 infection in an immature nervous system. *AIDS Res. Hum. Retroviruses*, **9**, 445–453.
66. WAHL S. M., ALLEN J. B., MCCARTNEY-FRANCIS N., MORGANTI-KOSSMANN M. C., KOSSMANN T., ELLINGSWORTH L., MAI U. E. H., MERGENHAGEN S. E. and ORENSTEIN J. M. (1991): Macrophage and astrocyte derived transforming growth factor- β as a mediator of central nervous system dysfunction in acquired immune deficiency syndrome. *J. Exp. Med.* **173**, 981–991.
67. WATKINS B. A., DORN H. H., KELLY W. B., ARMSTRONG R. C., POTTS B. J., MICHAELS F., KUFTA C. V. and DUBOIS-DALCQ M. (1990): Specific tropism of HIV-1 for microglial cells in primary human brain cultures. *Science*, **249**, 549–553.
68. WILEY C. A., BELMAN A. L., DICKSON D. W., RUBINSTEIN A. and NELSON J. A. (1990): Human immunodeficiency virus within the brains of children with AIDS. *Clin. Neuropathol.* **9**, 1–6.
69. WILEY C. A., SCHRIER R. D., NELSON J. A., LAMPERT P. W. and OLDSTONE M. B. A. (1986): Cellular localization of human immunodeficiency virus infection within the brains of acquired immune deficiency syndrome patients. *Proc. Natl. Acad. Sci., USA*, **83**, 7089–7093.
70. ZEICHNER S. L., HIRKA G., ANDREWS P. W. and ALWINE J. C. (1992): Differentiation-dependent human immunodeficiency virus long terminal repeat regulatory elements active in human teratocarcinoma cells. *J. Virol.* **66**, 2268–2273.
71. ZEICHNER S. L., KIM J. Y. H. and ALWINE J. C. (1991): Linker-scanning mutational analysis of the transcriptional activity of the human immunodeficiency virus type 1 long terminal repeats. *J. Virol.* **65**, 2436–2444.
72. ZHENG J., THYLIN M. R., GHORPADE A., XIONG H., PERSIDSKY Y., COTTER R., NIEMANN D., CHE M., ZENG Y. C., GELBARD H. A., SHEPARD R. B., SWARTZ J. M. and GENDELMAN H. E. (1999): Intracellular CXCR4 signaling, neuronal apoptosis and neuropathogenic mechanisms of HIV-1-associated dementia. *J. Neuroimmunol.* **98**, 185–200.

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