



Review

Type I Interferons as Immunoregulatory Molecules; Implications for Therapy in Experimental Autoimmune Uveoretinitis

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Abstract. Clinical trials have shown that the type I interferon (IFN)- α/β have some beneficial effects on organ-specific autoimmune diseases, such as Behcet's diseases and multiple sclerosis, although the precise mechanisms remain largely unresolved. T helper cells 1(Th1)-mediated autoimmune responses are involved in the initiation and/or progression of human uveitis, such as Behcet's disease. The animal model of experimental autoimmune uveoretinitis (EAU), characterized by a monophasic clinical course, has contributed to the understanding of the pathogenesis of human uveitis. Th1 producing IFN- γ induce EAU development, while Th2 producing IL-4/IL-10 prevent the disease. However, depending on the cytokine milieu, the pro-inflammatory cytokine IFN- γ may attenuate the autoimmune responses and anti-inflammatory cytokine IL-4 exacerbates it. Chemokines also play a crucial role in EAU development, which might be resolved by Th2-mediated immune responses. The administration of IFN- α/β prevents EAU development, accompanied by a diminished production of IFN- γ /IL-10. Interestingly, however, IFN- α/β also have some beneficial effects on patients with Th2-like phenotype in addition to Th1-like phenotypes. Thus, the immuno-modulatory action of IFN- α/β may be dependent on the context of cytokine combination and/or their concentrations.

Key words: interferon α/β ; autoimmune diseases; T helper cells; human uveitis.

Introduction

The development of an animal model of uveitis, experimental autoimmune uveitis (EAU)^{23, 77}, has contributed greatly to the understanding of the immuno-pathogenic mechanisms of human uveitis. Human uveitis, including Behcet's disease, sympathetic ophthalmia, and Vogt-Koyanagi-Harada disease, represents in-

traocular inflammatory diseases characterized by inflammatory attack of the uvea as well as the neuro-retina, resulting in severe vision loss and morbidity³⁶. EAU is predominantly induced by immunization with uveitogenic retinal antigens, S-antigens (S-Ag) or interphotoreceptor retinoid-binding protein (IRBP) in susceptible rats (Lewis)³⁶ and mice (at least H-2A^k)¹⁴.

Immune responses to uveitogenic antigens are in

Abbreviations used: EAU – experimental autoimmune uveoretinitis, IRBP – interphotoreceptor retinoid-binding protein, MHC – major histocompatibility complex, APC – antigen-presenting cell, Th – T helper, EAE – experimental autoimmune encephalomyelitis, IFN – interferon, DC – dendritic cell, mAb – monoclonal antibody, MS – multiple sclerosis.

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general not different from responses to foreign antigens such as pathogens or foods. T cells recognize fragments of antigens bound to molecules of the major histocompatibility complex (MHC) on antigen-presenting cells (APC), whereas B cells recognize the conformation-dependent antigenic structure. MHC comprises two types; class I and class II, which stimulate cytotoxic T lymphocytes (CTL) and T helper (Th) cells, respectively. Fragments generated in the cytosol of APC bind to class I molecules, while extracellular proteins endocytosed into the cytosol are processed and generally presented by MHC class II molecules to CD4-positive (CD4⁺) Th cells, exerting immuno-regulatory activities upon activation.

T cells, but not B cells, have been shown to play a predominant role in the pathogenesis of EAU^{12, 23, 77}. T cell-deficient mice, nu/nu or scid, are resistant to EAU development. Moreover, adoptive transfer of CD4⁺ T cells from mice primed with uveitogenic antigens or T cell clones specific for these antigens into naïve recipients induce the development of EAU, suggesting that CD4⁺ T cells are critical for disease development. Antigen-specific CD4⁺ T cells producing IFN- γ (type I (Th1) cells) play a critical role in disease development¹²², whereas those producing IL-4/IL-10 (Th2 cells) function in an antagonistic fashion^{58, 92}, as demonstrated in adoptive transfer experiments. Thus, the EAU model represents Th1-mediated organ-specific autoimmune diseases, which also include experimental autoimmune encephalomyelitis (EAE)⁷ and the non-obese diabetic mouse model of type I diabetes⁶³.

Type I interferons (IFNs) have been identified as virus-induced factors by virtue of their anti-viral activity and are classified into two groups by their physical properties, IFN- α and IFN- β ^{49, 95}. The IFN- α/β -induced beneficial effects in some organ-specific autoimmune diseases and tumors can be explained at least in part by the immuno-modulatory action of type I IFNs^{6, 110}. Although the precise mechanisms of IFN-mediated immune modulation remain largely unknown, they may involve activation of the innate as well as the adaptive immune systems. For example, IFN- α/β induce activation of natural killer (NK) cells and maturation and/or survival of dendritic cells (DCs)⁶⁴; these cells, together with macrophages, constitute the innate immunity.

The cells in the innate immune system display a rapid response upon antigen exposure and have a restricted number of receptors, called pattern recognition receptors (PRRs)^{1, 69}. These receptors, genetically predetermined, recognize highly conserved structures (pathogen-associated molecular patterns, PAMPS) present in a variety of micro-organisms. The recognition of the

PAMPS in mammals is mediated by Toll-like receptors (TLRs) expressed on APC such as DCs. The DCs are located everywhere in the body, and recognize certain invariant features, such as lipopolysaccharide (LPS), peptidoglycan, and CpG-rich DNA by the TLRs¹, triggering the production of cytokines, including type I IFNs. The activated DCs up-regulate MHC class I/class II and co-stimulatory molecules (CD80, CD86, and/or CD40) and, consequently, confer immunogenicity through delivery of the signals to T/B cells. Thus, the innate immune responses represent an efficient system, probably through the following processes: 1) removal of the pathogens, accompanied by recognition of the characteristics of the pathogens, and 2) transfer of the pathogen information to the adaptive immune system.

The T and B lymphocytes bear a diverse set of receptors for antigen, which are generated randomly during lymphocyte development. The lymphocytes bearing possibly useful receptors (specific for a foreign antigen) are positively selected through clonal expansion, whereas those reactive with self-antigens are deleted or aborted⁷⁶. If harmful lymphocytes with potential auto-reactivity were not deleted during development, they could play some role in the initiation of autoimmune diseases. The beneficial lymphocytes proliferate and differentiate into effector cells upon encounter with environmental antigens (in the context of MHC in case of T cells; adaptive immune systems). Since lymphocytes specific for an antigen are present in rather small number, they have to proliferate for efficient removal of the antigen. Although most of the lymphocytes are destined to die after removing antigens, a proportion of them survive, which enables them to proliferate and differentiate more rapidly upon re-encounter with the specific antigen (memory).

Considering the possible implication of viral infection in the pathogenesis of Behcet's disease and the anti-viral capacity of IFN- α , TSAMBAOS et al.¹²² used IFN- α to treat patients with Behcet's syndrome. A clinical trial of type I IFNs has demonstrated some benefits in certain patients with possible autoimmune-mediated inflammatory diseases including Behcet's disease, multiple sclerosis (MS), and inflammatory bowel diseases⁶, suggesting that IFNs have some immunomodulatory activities¹¹⁰. In addition, type I IFNs have anti-proliferative effects and have been used as therapies in some tumors, such as hairy cell leukemia, chronic myelogenous leukemia, myeloma, and squamous carcinoma^{43, 46}. In this article, we explore the pathogenesis of EAU as a model of organ-specific autoimmune diseases and elucidate the possible immu-

nomodulatory action of IFNs against organ-specific autoimmune diseases, including EAU.

Pathogenesis of EAU

Self-tolerance versus autoimmunity

The animal model of EAU has contributed greatly to the understanding of human uveitis, and autoimmune responses are somehow related to the initiation and/or progression of human uveitis. Since autoimmune responses are induced by a break-down of self tolerance, it is critical to understand how self tolerance is induced and maintained. The thymus plays a central role in the acquisition of self-tolerance (central tolerance or negative selection) through the interaction of thymic APCs with lymphocytes possibly having higher avidity to self antigens, resulting in the deletion of autoreactive lymphocytes⁷⁶. Conversely, lymphocytes with intermediate avidity to self antigens escape the selection in the thymus and migrate to peripheral tissues (positive selection), resulting in the formation of a T lymphocyte repertoire with different specificities to foreign antigens. For antigens derived from tissues (e.g. eye, brain, and testis) sequestered from the immune system, negative selection is thought to be incomplete because the immune system may not encounter these self antigens. However, a certain amount of uveitogenic antigens are reportedly expressed in the thymus²⁷ and pineal gland⁶⁷. It is possible that APCs fail to take up sufficient amounts of the autoantigens to trigger negative selection in the thymus, since lymphocytes (T and B) reacting with self antigens are found in normal circumstances in the peripheral lymphoid tissues. The observation that transgenic expression of uveitogenic antigen enhances self tolerance also suggests that sequestration contributes to immune privilege¹²³. Thus, potential uveitogenic Th1 cells might escape the selection and migrate to peripheral lymphoid organs. Indeed, these potential auto reactive lymphocytes are subsequently silenced in the peripheral organs (peripheral tolerance), probably through mechanisms such as the induction of anergy (functional paralysis), ignorance (failure to receive signals from APCs, resulting in cell death), and regulatory cells (active suppression of the possible autoreactive cells)⁹⁷. Thus, peripheral tolerance as well as sequestration of uveitogenic antigens from immunocompetent lymphocytes might compensate for the incomplete central tolerance.

What is the initial step in the activation of uveitogenic T cells? It is reasonable to assume that an event

such as trauma triggers the release of retinal antigens into the circulation, leading to activation of immune responses, since penetrative damage to one eye results in transient attack on the contralateral eye in sympathetic ophthalmia⁵⁰. The initial triggering of autoimmune responses may also be explained by the hypothesis that some pathogens (virus and bacteria) share the same T cell determinants with the host (molecular mimicry). Pathogen-specific T cells or infection may cause tissue damage resulting in the release of antigens. These antigens reach regional lymph nodes or the spleen, where the innate immunity is activated, further enhancing the autoimmune responses. For example, the cross-reaction between papilloma virus with brain antigens (myelin basic proteins) underlies encephalomyelitis in some patients with MS¹¹⁴.

The activation of uveitogenic T cells may result in the breakdown of intrinsic protective mechanisms (anterior chamber-associated immune deviation, ACAID) that guard against unfavorable inflammatory responses^{98, 121}. Intraocular APCs and soluble factors in the anterior chamber of the eye play a critical role in maintaining ACAID. For example, in contrast to conventional APCs, transforming growth factor (TGF)- β 2-treated intraocular APCs produce large amounts of mature TGF- β and diminished levels of IL-12. Consequently, in the presence of antigen, these APCs stimulate naïve T cells to generate substantial amounts of IL-4 and little IFN- γ , resulting in an anti-inflammatory condition^{101, 103}. In addition to TGF- β 2, the soluble factors include α melanocyte-stimulating hormone (α -MSH), vasoactive intestinal peptide (VIP), calcitonin gene-related peptide (CGRP), macrophage migration inhibitory factor (MIF), and IL-1 receptor antagonist^{2, 20, 108}. Moreover, ocular parenchymal cells expressing Fas-ligand (CD95-L) are considered to confer protection against possible invasion by activated CD95⁺ T cells through inducing apoptosis⁴⁵.

Th1 mediates EAU, while Th2 blocks it

CD4⁺ T cells play a critical role in the development of EAU. Indeed, oligoclonal expansion of CD4⁺ T cells is found in the inflamed sites in patients with Behcet's disease^{24, 32}. IRBP-specific T cell lines capable of inducing EAU predominantly express V β 8.3 gene products²⁶. CD4⁺ T cells are classified into two types by the pattern of cytokine production: Th1 cells produce IFN- γ , and Th2 cells produce IL-4, IL-5, and IL-10⁷¹. The IFN- γ produced by Th1 cells promotes further amplification of Th1 cells, while it prevents Th2 development. On the other hand, IL-4 promotes the develop-

ment of the Th2 phenotype and inhibits the Th1 phenotype. Adoptive transfer experiments have proved that the CD4⁺ cells with uveitogenic potential belong to the Th1 phenotype⁷⁷. The observations that transgenic expression of IFN- γ in rat retina following gene-mediated transfer accelerates the onset and exacerbates the severity of EAU^{28, 39} further support the notion that Th1 cells are critical for disease progression. Consistent with these findings, peripheral Th1 cells are significantly increased in patients with Behcet's disease³⁷. In contrast, the Th2 cytokine IL-10 ameliorates IRBP-induced EAU and even prevents this EAU when combined with IL-4⁹⁰. IL-10-deficient mice are more susceptible to EAU development. Kinetic study has demonstrated a predominant Th1-like responses in the draining lymph nodes of IRBP-immunized mice during the early phase, followed by Th2-type responses in the spleen during the resolution phase¹⁰⁴.

In addition to cytokines, the co-stimulatory molecules B7-1 (CD80) and B7-2 (CD86) expressed on APCs participate in some organ-specific autoimmune diseases by activating the development of Th1 and Th2 responses, respectively⁶². For example, IRBP-induced EAU development is prevented by anti-CD80 monoclonal antibody (mAb) pretreatment, whereas this EAU is substantially augmented by anti-CD86 mAb³⁸. Together, the balance between Th1 and Th2 appears to be critical for the development of EAU.

Th1 cytokine might exhibit different effects depending on the cytokine milieu

Several investigators have pointed out that systemic administration of the pro-inflammatory cytokine IFN- γ attenuates, and injection of IFN- γ -neutralizing mAb exacerbates Th1-mediated autoimmune diseases^{8, 13} including EAU. IFN- γ -deficient mice are also susceptible to development of both EAU and EAE with severity comparable to the wild types^{31, 53}, suggesting the redundancy of this cytokine in the induction of autoimmune responses. Interestingly, the inflamed eyes from IFN- γ -deficient mice display an allergic-like response in terms of inflammatory cell types and cytokine profile; infiltration of granulocytes and IL-5- and IL-6-producing cells, and elevated levels of IL-5, IL-6 and IL-10, with unaltered TNF- α ⁵³. Conversely, the Th2 cytokine IL-4 exacerbates Th1-mediated EAU in rats⁸⁸ and colitis in mice³⁵. Although the mechanisms underlying these apparently contradictory results remain unresolved, several factors may be assumed to contribute to the distinct phenomena; 1) genetic background of the animal model, 2) systemic versus local administration

of cytokines, and 3) dose and duration of the cytokine used. Activation and/or differentiation of T cells depend on the activation stage and/or subset of APCs, especially DCs, which constitute the innate immune system.

Role of innate immunity in EAU development

The innate immune system, comprising NK, phagocytic (macrophage, neutrophil), and DCs, displays rapid responses upon antigen exposure. DCs (professional APCs) are located in most tissues and have a prominent capacity to stimulate T and B cells^{64, 89}. Immature DCs first take up and process antigens and differentiate into mature DCs within a few days. The mature DCs then stimulate T cells. Infiltration of DCs into the lesion is found during EAU development in mouse and rat models^{48, 51}, suggesting a role of DCs in presenting potential uveitogenic antigens to T cells. Two distinct types of DC precursors, myeloid monocyte type (pre-DC1) and lymphoid type (pre-DC2)^{64, 87}, have been proposed as differentially influencing the balance of Th1/Th2 activation, although current data vary depending on animal species or maturation, isolation procedure, and tissue microenvironmental factors. These observations may reflect the functional plasticity of DCs depending on the context of their activation versus pathogen/tissues. Interestingly, myeloid DCs (CD8⁺CD11c⁺) appear at the time of disease onset and are required during IRBP-induced EAU in B10 mice⁵¹. These DCs may have some potential role in the induction or amplification of EAU development.

IL-12 secreted from DCs may represent a signal leading to Th1 differentiation of autoreactive T cells in several Th1-mediated autoimmune models, including EAU^{21, 94, 106, 124}. Genetic deletion of IL-12-p40 products by gene-targeting¹⁰⁶ or neutralization of IL-12 by mAb¹²⁴ attenuates the induction of uveitis in mice following IRBP stimulation. The diminished EAU-inducing capacity of the IRBP-primed lymphocytes from wild-type mice upon transfer to IL-12-deficient mice suggests that IL-12 also functions in the efferent loop of EAU development. Although IL-12 and IL-18 in combination promote IFN- γ production, resulting in NK activation⁷³, the role of IL-18 in EAU development remains obscure⁵².

In contrast to the pro-inflammatory features described above, administration of recombinant IL-12 ameliorates EAU induction, accompanied by high serum levels of IFN- γ and enhanced apoptosis in the draining lymph node¹⁰⁷. The elevated IFN- γ level activates iNOS, resulting in NO production, followed by down-regulation of the anti-apoptotic protein Bcl-2. It

may also be possible that IFN- γ up-regulates the death-inducing systems, CD95-L/CD95⁵⁶, TNF-related apoptosis-inducing ligand (TRAIL)/TRAIL-receptors (TRAIL-Rs)⁴⁴, and/or TNF-like weak inducer of apoptosis (TWEAK)/DR-3⁷⁴. In summary, IL-12 appears to mediate a variety of effects on the pathogenesis of Th1-mediated EAU, depending on the phase of the immune response and the cell subset with which they react.

Participation of chemokines in the recruitment of inflammatory cells into the eye

Following the initiation of specific immune responses against uveitogenic antigens in local regional lymph nodes, recruitment of non-specific inflammatory cells into the eyes is required for the development of EAU⁸⁶. Cell migration involves chemokines, which are classified into two categories in terms of their physiologic features; homeostatic (necessary for the maintenance of physiologic traffic) and inducible chemokines⁷⁰. The inducible chemokines are substantially up-regulated upon inflammatory stimulation in a variety of cells and likely participate in the development of inflammatory diseases, including autoimmune diseases⁴⁰. Chemokines such as monocyte chemoattractant protein 1 (MCP-1), regulated on activation and normal T cell expressed and secreted (RANTES), and macrophage inflammatory protein 1 (MIP-1) α/β are expressed in the eye during EAU development, with MIP-1 α already detected before clinical disease onset^{18, 117}. Mice with IRBP-induced ACAID are protected from EAU, accompanied by diminished expressions of RANTES and MCP-1 mRNA (TAKEUCHI and USUI et al., submitted for publication). The participation of adhesion molecules and chemokines is further demonstrated by the blockade of EAU by mAbs to intercellular adhesion molecule-1 (CD54; ICAM-1 on epithelial cells), leukocyte function-associated antigen-1 (CD11a/CD18; LFA-1 on lymphocytes)^{113, 120}, and macrophage MIF⁵⁹. Treatment with pertussis toxin inhibits functional coupling of the receptor to Gi protein and ameliorates EAU development, probably through preventing migration of effector cells into the eye lesion⁹⁹.

Effector mechanisms in EAU development

CD4⁺ T cells play a crucial role in tissue damage in Th1-mediated organ-specific autoimmune diseases¹¹⁸. Tissue damage is mediated in part through apoptotic cell death. Apoptotic cell death is regulated by at least two components; members of the TNF receptor family (CD95, TRAIL-Rs, and DR-3/TRAMP) activate caspase-8, and members of the Bcl-2 family

regulate the release of cytochrome c from mitochondria to activate caspase-3. Apoptosis has been demonstrated in the uveitic eyes of patients with Behcet's disease¹⁶ and rats with S-Ag-induced EAU¹⁰⁰, which appears to involve the CD95-L/CD95 pathway. Mice homozygous for the mutant genes *lpr* and *gld* are deficient in functional CD95 and CD95-L, respectively, and spontaneously develop progressive autoimmune symptoms. Interestingly, these mice are resistant to induction of EAE¹¹⁸ and EAU¹¹⁶. CD95-L on immuno-compotent cells and CD95 on target tissues are necessary for the development of EAU. Moreover, the CD95-independent pathway is also suggested to be involved in the tissue damage of EAE²⁵. The classical pro-inflammatory pathway, characterized by secretion of the inflammatory cytokines TNF- α , IL-1, and NO, is activated in macrophages, resulting in development of EAU^{34, 109}. Conversely, apoptosis may also limit the damage caused by autoimmune responses. Expression of CD95-L in the anterior chamber of the eye is thought to contribute to the maintenance of the immune-privileged status through inducing apoptosis of possible invading CD95⁺ lymphocytes⁴⁵. CD95-L expressed on host cells is required for remission of EAE¹¹⁶.

Resolution of EAU

The clinical course of EAU is monophasic, characterized by activation and then down-modulation of autoimmune responses. Determinant spreading may represent the mechanism underlying the resolution of EAU, although other mechanisms may also be involved. Determinant spreading refers to the notion that immune responses initiated from a single determinant expand to other determinants. Immunization with a certain portion of IRBP causes diseases with the dominant Th1 responses, whereas immunization with other portions prevents diseases accompanied by Th2 responses^{58, 102}, suggesting that T cells reactive to spreading determinants may either protect or provoke diseases depending on the background cytokine profile. Based on the finding that the course of EAU is monophasic, it may be possible to assume that determinant spreading to Th2 polarization contributes to the resolution of the disease. Indeed, KEINO et al.⁵⁷ have reported clonal expansion of p518-529-reactive Th2-like cells in the uveitic eyes during the late resolution stage of EAU.

Action of Type I IFN

IFNs have immuno-modulatory effects *in vitro* and *in vivo*, which might in part account for the IFN-medi-

ated amelioration of some autoimmune diseases such as Behcet's diseases and MS. The immuno-modulatory effects likely involve an intricate balance among the Th subsets (Th1, Th2, and T regulatory cells)¹¹⁰, activation of DC or NK cells, and migration of inflammatory cells⁹. Since cell proliferation is required for immune responses, the anti-proliferative as well as pro-apoptotic effects of IFNs^{30, 44, 83, 105} may be somehow implicated in the IFN-mediated improvement in patients with Behcet's diseases. For example, IFN- α induces TRAIL expression in monocytes, resulting in a limitation of immune responses through the induction of apoptosis^{30, 44}. The blockade of virus replication by IFNs⁴ may also be beneficial for diseases involving virus-triggered autoimmune responses.

Type I IFNs appear to favor Th1 differentiation as well as Th1-type responses^{22, 84}. For example, priming of human neonatal CD4⁺ T cells with IFN- α plus anti-CD3 mAb promotes Th1 development through increasing both IFN- γ and IL-10 and impairing IL-4 production, although IFN- α alone is not sufficient to induce the differentiation. The expression of IL-12R (β 2), consisting of β 1 and β 2 chains, is also up-regulated by IFNs in human T cell lines⁴². Moreover, IFNs elicit IL-15 mRNA in DCs^{91, 125}, and IL-15 synergizes with IL-12 for IFN- γ production³, favoring development of the Th1 phenotype.

DC precursor 2 exposed to virus produces a large amount of IFNs¹⁵ that function as survival and maturation factors^{65, 66}. The virus-activated mature DCs stimulate naïve CD4⁺ cells to produce IFN- γ and IL-10, while IL-3-induced DCs produce the Th2-type cytokines IL-4 and IL-10⁵⁴, suggesting that DCs play a critical role in linking innate and adaptive immunity. Since a T cell subset that produces IL-10 and IFN- γ is found during some infections^{41, 93}, it is interesting to check whether this subset may contribute to uveitogenic antigen-induced IL-10/IFN- γ production during EAU progression^{5, 80}.

In contrast to the pro-inflammatory properties, recent reports suggest that IFNs negatively regulate IL-12 production by DCs dependently¹¹⁹ or independently⁶⁸ of IL-10. A greater amount of IFNs appears to be required for the anti-inflammatory effects compared with the pro-inflammatory actions. Although IL-12 is induced by APCs during bacterial and parasite infections⁴⁷, and promotes NK and Th1 cell IFN- γ production, biologically active IL-12 is not induced during viral infections, including lymphocyte choriomeningitis virus (LCMV). Rather, a variety of viruses elicit a large amount of IFNs with enhanced expansion of CD8⁺ killer cells¹⁷. Although the CD8⁺ IFN- γ production is dependent on

IFNs, substitution of IL-12 occurs, leading to IFN- γ production in the absence of IFNs, suggesting the plasticity of the cytokine to achieve a similar goal through different activation pathways. Thus, IFNs are pleiotropic cytokines with a variety of activities on many cell types, including T cells, DCs, and NK cells.

IFN- α/β -Mediated Amelioration on EAU

Clinical trials done on a small scale have provided evidence of some benefits of IFNs in suppressing inflammation and reducing recurrence in patients with Th1-mediated uveitis associated with Behcet's disease^{33, 61, 78}. IFN- α has also been demonstrated to be effective against diseases with other Th1-like and also Th2-like phenotypes⁶. The mechanisms by which IFNs confer beneficial effects in a variety of diseases remain unresolved. Animal models using rats and mice have contributed to some extent to the understanding of the efficacy of IFN- α/β against autoimmune diseases, including EAU^{55, 79, 80}. For example, administration of IFN- α/β suppresses EAU induced by IRBP in complete Freund's adjuvant^{79, 80}. Lewis rats administered with IFN- α/β during the afferent phase display decreased intraocular inflammation after IRBP immunization. On the other hand, IFN- α/β provide no protection when administered during the efferent phase, suggesting that IFNs somehow affect the initiation step of EAU development. In IRBP-stimulated spleen cells, IFN- α mediates inhibition of TNF- α production, but does not alter the production of IFN- γ , IL-4, and IL-10. In addition to Th1-/Th2-like cytokines, TNF- α has been shown to be involved in EAU development⁷². There are also many studies that support the critical involvement of TNF- α in other autoimmune diseases, including rheumatoid arthritis⁶⁰ and EAE⁸⁵.

Based on the findings that several types of inflammatory cells accumulate in intraocular lesion during EAU development, it is reasonable to assume that the types and concentrations of cytokines in the retina as well as in systemic lymphoid organs play a critical role in disease progression. The kinetics of IFN- γ and also anti-inflammatory cytokine IL-10 production in intraocular extracts from Lewis rats roughly correlates with the development of IRBP-induced EAU: it is undetectable before onset of inflammation, markedly elevated at peak inflammation, and undetectable again at resolution⁸¹. A similar pattern of IFN- γ /IL-10 mRNA production is induced by S-Ag⁵, although the increase in IL-10 mRNA is slower than when induced by IRBP. Concomitant production of IL-10 in EAU development

might account for the non-relapsing and non-chronic feature of the EAU model. IFN- α/β substantially suppress production of IFN- γ /IL-10 induced by IRBP, while slightly enhancing the IL-4 level. The considerable down-regulation of the cytokine profile mediated by IFN- α/β is not compatible with the slight suppression of anterior segment intraocular inflammation as assessed by biomicroscope. These findings support the notion that the cytokine requirement is redundant for the development of Th1-mediated autoimmune diseases, including EAU. In addition to IL-12, several candidates have been reported to trigger IFN- γ production, including IL-18 (IFN- γ -inducing factor, IGIF)⁷³ and the recently identified IL-12-like cytokine IL-23⁸², composed of IL-12-p40 and p19. Alternatively, anterior chamber inflammation may be somewhat different from posterior uveitis, and the latter is more sensitive to IFN- α/β treatment.

Several investigators have reported the beneficial effects of oral administration of low-dose IFNs for a variety of diseases including autoimmune-mediated diseases^{19, 111}. Long-term (3 consecutive weeks) oral administration of IFN- β to Lewis rats inhibited IRBP-induced EAU, accompanied by partial inhibition of IRBP-induced spleen cell proliferation and IFN- γ production (SUZUKI et al., submitted for publication). Oral feeding of type I IFNs has also been shown to be effective against relapsing EAE in mice^{10, 75}. Interestingly, oral administration of MBP and IFN- β suppresses EAE in a synergistic fashion⁷⁵. Since the blood levels of IFN- α/β after oral administration are below the detection limit²⁹, it is possible that some immunoregulatory cells generated in mucosal lymph nodes somehow affect immune responses in the spleen, as suggested by the EAE model¹¹.

In spite of the beneficial effects of IFN- α/β on Th1-mediated EAU and MS, clinical reports of side effects have shown that IFN- α/β have pro-inflammatory activities *in vivo*; IFN- α/β treatment is linked with the exacerbation or development of several types of autoimmune diseases, including insulin-dependent diabetes mellitus (IDDM) and thyroiditis^{110, 115}. Transgenic expression of IFN- α by insulin-producing β cells has been shown to cause type I diabetes in mice⁹⁶. On the other hand, hypereosinophilic syndrome and discoid lupus erythematosus, characterized by Th2 responses, are ameliorated to some extent by IFN- α/β ^{6, 46}. As described above, the nature of IFN- α/β as pro-inflammatory cytokines might counteract Th2-mediated diseases, alleviating them. At present, it remains largely unresolved how IFN- α/β exert beneficial effects on both Th1- and Th2-mediated autoimmune diseases.

Cytokines appear to stimulate Th1-like or Th2-like responses, depending on their concentrations, exposure time, and the cytokine milieu. For example, IL-18 combined with IL-12 induces high IFN- γ production (Th1 response) in naïve T cells, whereas IL-18 and IL-2 together induce IL-4 production (Th2 response)⁷³. Upon exposure to intracellular bacteria (low IFN producer), only a small amount of IFN- α/β is produced, and this IFN- α/β in conjunction with IL-12 enhances IFN- γ production by CD4⁺ and CD8⁺ cells, promoting Th1-type responses. In some viral infections where a large amount of IFN- α/β may be produced, high levels of IFN- α/β inhibit IL-12 production and NK cell IFN- γ production, favoring the Th2-phenotype. Concurrent amplification of CD8⁺ T cell responses with the help of IL-15 contributes to resistance to viral infection. Thus, the IFN-mediated beneficial effects on organ-specific diseases may be dependent on the context of cytokine combination and/or their concentrations.

Conclusions and Perspectives

Human uveitis such as Behcet's disease is thought to involve Th1-mediated immune responses for the initiation and/or progression of the disease, although we do not have enough data to prove it. The animal EAU model has contributed in part to the understanding of the pathogenesis of the human uveitis. CD4⁺ Th1 cells play a critical role in the initiation and/or progression of EAU. Recently, the innate immune system, including DCs, has been demonstrated to play a central role in the initiation of autoimmune responses by capturing antigen, processing it, and delivering the pathogen information to lymphocytes in the adaptive immune system. It is interesting how DC activation or IL-12 activates the uveitogenic CD4⁺ Th1 cells. Type I IFNs have some beneficial effects on organ-specific autoimmune diseases, including EAE and EAU, although the mechanisms underlying the IFN- α/β -mediated effects remain obscure. Assuming that autoimmune processes are somehow implicated in the progression of Behcet's disease, several explanations may be proposed: 1) IFNs shift the balance of Th1-/Th2-phenotype at the level of DCs or T cells; 2) IFNs interfere with the up-regulation of chemokines and/or adhesion molecules, resulting in diminished recruitment of inflammatory cells into the lesion; 3) IFNs limit the clonal expansion of the T cells specific for autoantigens in autoimmune diseases, including EAU, probably through their anti-proliferative effects; or 4) IFN- α/β -mediated anti-viral effects inhibit the initial and/or amplification processes by which

autoimmune diseases, including EAU, are exacerbated. Although much has to be done to verify these possibilities an understanding of IFN actions would hopefully open the way to a novel strategy for the IFN- α/β -mediated treatment modality, with great benefits to patients with autoimmune diseases such as Behcet's diseases and MS.

Acknowledgment. We thank Prof. J. Patrick Barron (International Medical Communications Center, Tokyo Medical University) for reading of the manuscript.

References

- ADEREM A. and ULEVITCH R. J. (2000): Toll-like receptors in the induction of the innate immune response. *Nature*, **406**, 782–787.
- APTE R. S., SINHA D., MAYHEW E., WISTOW G. J. and NIEDERKORN J. Y. (1998): Cutting edge: role of macrophage migration inhibitory factor in inhibiting NK cell activity and preserving immune privilege. *J. Immunol.*, **160**, 5693–5696.
- AVICE M. N., DEMEURE C. E., DELESPESE G., RUBIO M., ARMANT M. and SARFATI M. (1998): IL-15 promotes IL-12 production by human monocytes via T cell-dependent contact and may contribute to IL-12-mediated IFN- γ secretion by CD4⁺ T cells in the absence of TCR ligation. *J. Immunol.*, **161**, 3408–3415.
- BARBER G. N. (2001): Host defense, viruses and apoptosis. *Cell Death Differ.*, **8**, 113–126.
- BARTON K., MCLAUCHLAN M. T., CALDER V. L. and LIGHTMAN S. (1995): The kinetics of cytokine mRNA expression in the retina during experimental autoimmune uveoretinitis. *Cell. Immunol.*, **164**, 133–140.
- BELARDELLI F. and GRESSER I. (1996): The neglected role of type I interferon in the T-cell response: implications for its clinical use. *Immunol. Today*, **17**, 369–372.
- BETTELLI E. and NICHOLSON L. B. (2000): The role of cytokines in experimental autoimmune encephalomyelitis. *Arch. Immunol. Ther. Exp.*, **48**, 389–398.
- BILLIAU A., HEREMANS H., VANDEKERCKHOVE F., DIJKMANS R., SOBIS H., MEULEPAS E. and CARTON H. (1988): Enhancement of experimental allergic encephalomyelitis in mice by antibodies against IFN- γ . *J. Immunol.*, **140**, 1506–1510.
- BIRON C. A. (2001): Interferons alpha and beta as immune regulators – a new look. *Immunity*, **14**, 661–664.
- BROD S. A. and BURNS D. K. (1994): Suppression of relapsing experimental autoimmune encephalomyelitis in the SJL/J mouse by oral administration of type I interferons. *Neurology*, **44**, 1144–1148.
- BROD S. A., KHAN M., KERMAN R. H. and PAPPOLLA M. (1995): Oral administration of human or murine interferon alpha suppresses relapses and modifies adoptive transfer in experimental autoimmune encephalomyelitis. *J. Neuroimmunol.*, **58**, 61–69.
- CASPI R. R. (1999): Immune mechanisms in uveitis. *Springer Semin. Immunopathol.*, **21**, 113–124.
- CASPI R. R., CHAN C. C., GRUBBS B. G., SILVER P. B., WIGGERT B., PARSA C. F., BAHMANYAR S., BILLIAU A. and HEREMANS H. (1994): Endogenous systemic IFN- γ has a protective role against ocular autoimmunity in mice. *J. Immunol.*, **152**, 890–899.
- CASPI R. R., GRUBBS B. G., CHAN C. C., CHADER G. J. and WIGGERT B. (1992): Genetic control of susceptibility to experimental autoimmune uveoretinitis in the mouse model. Concomitant regulation by MHC and non-MHC genes. *J. Immunol.*, **148**, 2384–2389.
- CELLA M., JARROSSAY D., FACCHETTI F., ALEBARDI O., NAKAJIMA H., LANZAVECCHIA A. and COLONNA M. (1999): Plasmacytoid monocytes migrate to inflamed lymph nodes and produce large amounts of type I interferon. *Nat. Med.*, **5**, 919–923.
- CHAN C. C., MATTESON D. M., LI Q., WHITCUP S. M. and NUSSENBLATT R. B. (1997): Apoptosis in patients with posterior uveitis. *Arch. Ophthalmol.*, **115**, 1559–1567.
- COUSENS L. P., PETERSON R., HSU S., DORNER A., ALTMAN J. D., AHMED R. and BIRON C. A. (1999): Two roads diverged: interferon alpha/beta- and interleukin 12-mediated pathways in promoting T cell interferon gamma responses during viral infection. *J. Exp. Med.*, **189**, 1315–1328.
- CRANE I. J., MCKILLOP-SMITH S., WALLACE C. A., LAMONT G. R. and FORRESTER J. V. (2001): Expression of the chemokines MIP-1 α , MCP-1, and RANTES in experimental autoimmune uveitis. *Invest. Ophthalmol. Vis. Sci.*, **42**, 1547–1552.
- CUMMINS J. M., BEILHARZ M. W. and KRAKOWKA S. (1999): Oral use of interferon. *J. Interferon Cytokine Res.*, **19**, 853–857.
- DANA M. R., DAI R., ZHU S., YAMADA J. and STREILEIN J. W. (1998): Interleukin-1 receptor antagonist suppresses Langerhans cell activity and promotes ocular immune privilege. *Invest. Ophthalmol. Vis. Sci.*, **39**, 70–77.
- DAVIDSON N. J., HUDAK S. A., LESLEY R. E., MENON S., LEACH M. W. and RENNICK D. M. (1998): IL-12, but not IFN- γ , plays a major role in sustaining the chronic phase of colitis in IL-10-deficient mice. *J. Immunol.*, **161**, 3143–3149.
- DEMEURE C. E., WU C. Y., SHU U., SCHNEIDER P. V., HEUSSER C., YSSEL H. and DELESPESE G. (1994): *In vitro* maturation of human neonatal CD4 T lymphocytes. II. Cytokines present at priming modulate the development of lymphokine production. *J. Immunol.*, **152**, 4775–4782.
- DICK A. D. (2000): Immune mechanisms of uveitis: insights into disease pathogenesis and treatment. *Int. Ophthalmol. Clin.*, **40**, 1–18.
- DIRESKENELI H., EKSIÖGLU-DEMIRALP E., KIBAROGLU A., YAVUZ S., ERGUN T. and AKOGLU T. (1999): Oligoclonal T cell expansions in patients with Behcet's disease. *Clin. Exp. Immunol.*, **117**, 166–170.
- DITTEL B. N., MERCHANT R. M. and JANEWAY C. A. JR. (1999): Evidence for Fas-dependent and Fas-independent mechanisms in the pathogenesis of experimental autoimmune encephalomyelitis. *J. Immunol.*, **162**, 6392–6400.
- EGWUAGU C. E., BAHMANYAR S., MAHDI R. M., NUSSENBLATT R. B., GERY I. and CASPI R. R. (1992): Predominant usage of V beta 8.3 T cell receptor in a T cell line that induces experimental autoimmune uveoretinitis (EAU). *Clin. Immunol. Immunopathol.*, **65**, 152–160.
- EGWUAGU C. E., CHARUKAMNOETKANOK P. and GERY I. (1997): Thymic expression of autoantigens correlates with resistance to autoimmune disease. *J. Immunol.*, **159**, 3109–3112.
- EGWUAGU C. E., SZTEIN J., MAHDI R. M., LI W., CHAO-CHAN C., SMITH J. A., CHARUKAMNOETKANOK P. and CHEPELINSKY A. B. (1999): IFN- γ increases the severity and accelerates the

- onset of experimental autoimmune uveitis in transgenic rats. *J. Immunol.*, **162**, 510–517.
29. EID P., MERITET J. F., MAURY C., LASFAR A., WEILL D. and TOVEY M. G. (1999): Oromucosal interferon therapy: pharmacokinetics and pharmacodynamics. *J. Interferon Cytokine Res.*, **19**, 157–169.
 30. FANGER N. A., MALISZEWSKI C. R., SCHOOLEY K. and GRIFFITH T. S. (1999): Human dendritic cells mediate cellular apoptosis via tumor necrosis factor-related apoptosis-inducing ligand (TRAIL). *J. Exp. Med.*, **190**, 1155–1164.
 31. FERBER I. A., BROCKE S., TAYLOR-EDWARDS C., RIDGWAY W., DINISCO C., STEINMAN L., DALTON D. and FATHMAN C. G. (1996): Mice with a disrupted IFN-gamma gene are susceptible to the induction of experimental autoimmune encephalomyelitis (EAE). *J. Immunol.*, **156**, 5–7.
 32. FERON E. J., CALDER V. L. and LIGHTMAN S. L. (1995): Oligoclonal activation of CD4⁺ T lymphocytes in posterior uveitis. *Clin. Exp. Immunol.*, **99**, 412–418.
 33. FERON E. J., ROTHOVA A., VAN HAGEN P. M., BAARSMA G. S. and SUTTORP-SCHULTEN M. S. (1994): Interferon-alpha 2b for refractory ocular Behcet's disease. *Lancet*, **343**, 1428.
 34. FORRESTER J. V., HUITINGA I., LUMSDEN L. and DIJKSTRA C. D. (1998): Marrow-derived activated macrophages are required during the effector phase of experimental autoimmune uveoretinitis in rats. *Curr. Eye Res.*, **17**, 426–437.
 35. FORT M., LESLEY R., DAVIDSON N., MENON S., BROMBACHER F., LEACH M. and RENNICK D. (2001): IL-4 exacerbates disease in a Th1 cell transfer model of colitis. *J. Immunol.*, **166**, 2793–2800.
 36. FOX G. M., KUWABARA T., WIGGERT B., REDMOND T. M., HESS H. H., CHADER G. J. and GERY I. (1987): Experimental autoimmune uveoretinitis (EAU) induced by retinal interphotoreceptor retinoid-binding protein (IRBP): differences between EAU induced by IRBP and by S-antigen. *Clin. Immunol. Immunopathol.*, **43**, 256–264.
 37. FRASSANITO M. A., DAMMACO R., CAFFORIO P. and DAMMACO F. (1999): Th1 polarization of the immune response in Behcet's disease: a putative pathogenetic role of interleukin-12. *Arthritis Rheum.*, **42**, 1967–1974.
 38. FUKAI T., OKADA A. A., SAKAI J., KEZUKA T., KEINO H., USUI M., YAGITA H., OKUMURA K. and MIZUGUCHI J. (1999): The role of costimulatory molecules B7-1 and B7-2 in mice with experimental autoimmune uveoretinitis. *Graefes Arch. Clin. Exp. Ophthalmol.*, **237**, 928–933.
 39. GEIGER K., HOWES E., GALLINA M., HUANG X. J., TRAVIS G. H. and SARVETNICK N. (1994): Transgenic mice expressing IFN-gamma in the retina develop inflammation of the eye and photoreceptor loss. *Invest. Ophthalmol. Vis. Sci.*, **35**, 2667–2681.
 40. GERARD C. and ROLLINS B. J. (2001): Chemokines and disease. *Nat. Immunol.*, **2**, 108–115.
 41. GEROSA F., NISII C., RIGHETTI S., MICCIOLO R., MARCHESINI M., CAZZADORI A. and TRINCHIERI G. (1999): CD4(+) T cell clones producing both interferon-gamma and interleukin-10 predominate in bronchoalveolar lavages of active pulmonary tuberculosis patients. *Clin. Immunol.*, **92**, 224–234.
 42. GOLLOB J. A., KAWASAKI H. and RITZ J. (1997): Interferon-gamma and interleukin-4 regulate T cell interleukin-12 responsiveness through the differential modulation of high-affinity interleukin-12 receptor expression. *Eur. J. Immunol.*, **27**, 647–652.
 43. GRESSER I. (1997): Wherefore interferon? *J. Leukoc. Biol.*, **61**, 567–574.
 44. GRIFFITH T. S., WILEY S. R., KUBIN M. Z., SEDGER L. M., MALISZEWSKI C. R. and FANGER N. A. (1999): Monocyte-mediated tumoricidal activity via the tumor necrosis factor-related cytokine, TRAIL. *J. Exp. Med.*, **189**, 1343–1354.
 45. GRIFFITH T. S., YU X., HERNDON J. M., GREEN D. R. and FERGUSON T. A. (1996): CD95-induced apoptosis of lymphocytes in an immune privileged site induces immunological tolerance. *Immunity*, **5**, 7–16.
 46. GUTTERMAN J. U. (1994): Cytokine therapeutics: lessons from interferon alpha. *Proc. Natl. Acad. Sci. USA*, **91**, 1198–1205.
 47. HEINZEL F. P., RERKO R. M., LING P., HAKIMI J. and SCHOENHAUT D. S. (1994): Interleukin 12 is produced *in vivo* during endotoxemia and stimulates synthesis of gamma interferon. *Infect. Immun.*, **62**, 4244–4249.
 48. ISHIMOTO S., ZHANG J., GULLAPALLI V. K., PARARAJASEGARAM G. and RAO N. A. (1998): Antigen-presenting cells in experimental autoimmune uveoretinitis. *Exp. Eye Res.*, **67**, 539–548.
 49. ISSACS A. and LINDEMANN J. (1957): Virus interference. I. The interferon. *Proc. R. Acad. Britain*, **147**, 258.
 50. JAKOBIEC F. A., LEFKOWITZ J. and KNOWLES D. M. 2ND. (1984): B- and T-lymphocytes in ocular disease. *Ophthalmology*, **91**, 635–654.
 51. JIANG H. R., LUMSDEN L. and FORRESTER J. V. (1999): Macrophages and dendritic cells in IRBP-induced experimental autoimmune uveoretinitis in B10RIII mice. *Invest. Ophthalmol. Vis. Sci.*, **40**, 3177–3185.
 52. JIANG H. R., WEI X., NIEBALA W., LUMSDEN L., LIEW F. Y. and FORRESTER J. V. (2001): IL-18 not required for IRBP peptide-induced EAU: studies in gene-deficient mice. *Invest. Ophthalmol. Vis. Sci.*, **42**, 177–182.
 53. JONES L. S., RIZZO L. V., AGARWAL R. K., TARRANT T. K., CHAN C. C., WIGGERT B. and CASPI R. R. (1997): IFN-gamma-deficient mice develop experimental autoimmune uveitis in the context of a deviant effector response. *J. Immunol.*, **158**, 5997–6005.
 54. KADOWAKI N., ANTONENKO S., LAU J. Y. and LIU Y. J. (2000): Natural interferon alpha/beta-producing cells link innate and adaptive immunity. *J. Exp. Med.*, **192**, 219–226.
 55. KARP C. L., BIRON C. A. and IRANI D. N. (2000): Interferon beta in multiple sclerosis: is IL-12 suppression the key? *Immunol. Today*, **21**, 24–28.
 56. KASER A., NAGATA S. and TILG H. (1999): Interferon alpha augments activation-induced T cell death by upregulation of Fas (CD95/APO-1) and Fas ligand expression. *Cytokine*, **11**, 736–743.
 57. KEINO H., TAKEUCHI M., SUZUKI J., KOJO S., SAKAI J., NISHIOKA K., SUMIDA T. and USUI M. (2001): Identification of Th2-type suppressor T cells among *in vivo* expanded ocular T cells in mice with experimental autoimmune uveoretinitis. *Clin. Exp. Immunol.*, **124**, 1–8.
 58. KEZUKA T., SAKAI J., YOKOI H., TAKEUCHI M., OKADA A., TAGUCHI O., USUI M. and MIZUGUCHI J. (1996): Peptide-mediated suppression of experimental autoimmune uveoretinitis in mice: development of a peptide vaccine. *Int. Immunol.*, **8**, 1229–1235.
 59. KITAICHI N., MATSUDA A., KOTAKE S., NAMBA K., TAGAWA Y., SASAMOTO Y., OGASAWARA K., IWABUCHI K., ONOE K., MATSUDA H. and NISHIHARA J. (2000): Inhibition of experimental autoimmune uveoretinitis with anti-macrophage migration inhibitory factor antibodies. *Curr. Eye Res.*, **20**, 109–114.
 60. KLARESKOG L. and MCDEVITT H. (1999): Rheumatoid arthritis

- and its animal models: the role of TNF- α and the possible absence of specific immune reactions. *Curr. Opin. Immunol.*, **11**, 657–662.
61. KOTTER I., ECKSTEIN A. K., STUBIGER N. and ZIERHUT M. (1998): Treatment of ocular symptoms of Behcet's disease with interferon α 2a: a pilot study. *Br. J. Ophthalmol.*, **82**, 488–494.
 62. KUCHROO V. K., DAS M. P., BROWN J. A., RANGER A. M., ZAMVIL S. S., SOBEL R. A., WEINER H. L., NABAVI N. and GLIMCHER L. H. (1995): B7-1 and B7-2 costimulatory molecules activate differentially the Th1/Th2 developmental pathways: application to autoimmune disease therapy. *Cell*, **80**, 707–718.
 63. LIBLAU R. S., SINGER S. M. and MCDEVITT H. O. (1995): Th1 and Th2 CD4⁺ T cells in the pathogenesis of organ-specific autoimmune diseases. *Immunol. Today*, **16**, 34–38.
 64. LIU Y. J., KANZLER H., SOUMELIS V. and GILLIET M. (2001): Dendritic cell lineage, plasticity and cross-regulation. *Nat. Immunol.*, **2**, 585–589.
 65. LUFT T., PANG K. C., THOMAS E., HERTZOG P., HART D. N., TRAPANI J. and CEBON J. (1998): Type I IFNs enhance the terminal differentiation of dendritic cells. *J. Immunol.*, **161**, 1947–1953.
 66. MARRACK P., KAPPLER J. and MITCHELL T. (1999): Type I interferons keep activated T cells alive. *J. Exp. Med.*, **189**, 521–530.
 67. MCALLISTER C. G., WIGGERT B., CHADER G. J., KUWABARA T. and GERY I. (1987): Uveitogenic potential of lymphocytes sensitized to interphotoreceptor retinoid-binding protein. *J. Immunol.*, **138**, 1416–1420.
 68. MCRAE B. L., SEMNANI R. T., HAYES M. P. and VAN SEVENTER G. A. (1998): Type I IFNs inhibit human dendritic cell IL-12 production and Th1 cell development. *J. Immunol.*, **160**, 4298–4304.
 69. MEDZHITOV R. and JANEWAY C., JR. (2000): Innate immunity. *N. Engl. J. Med.*, **343**, 338–344.
 70. MOSER B. and LOETSCHER P. (2001): Lymphocyte traffic control by chemokines. *Nat. Immunol.*, **2**, 123–128.
 71. MOSMANN T. R. and COFFMAN R. L. (1989): TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. *Annu. Rev. Immunol.*, **7**, 145–173.
 72. NAKAMURA S., YAMAKAWA T., SUGITA M., KIJIMA M., ISHIOKA M., TANAKA S. and OHNO S. (1994): The role of tumor necrosis factor- α in the induction of experimental autoimmune uveoretinitis in mice. *Invest. Ophthalmol. Vis. Sci.*, **35**, 3884–3889.
 73. NAKANISHI K., YOSHIMOTO T., TSUTSUI H. and OKAMURA H. (2001): Interleukin-18 regulates both Th1 and Th2 responses. *Annu. Rev. Immunol.*, **19**, 423–474.
 74. NAKAYAMA M., KAYAGAKI N., YAMAGUCHI N., OKUMURA K. and YAGITA H. (2000): Involvement of TWEAK in interferon gamma-stimulated monocyte cytotoxicity. *J. Exp. Med.*, **192**, 1373–1380.
 75. NELSON P. A., AKSELBAND Y., DEARBORN S. M., AL-SABBAGH A., TIAN Z. J., GONNELLA P. A., ZAMVIL S. S., CHEN Y. and WEINER H. L. (1996): Effect of oral beta interferon on subsequent immune responsiveness. *Ann. N. Y. Acad. Sci.*, **778**, 145–155.
 76. NOSSAL G. J. (1994): Negative selection of lymphocytes. *Cell*, **76**, 229–239.
 77. NUSSENBLATT R. B. and GERY I. (1996): Experimental autoimmune uveitis and its relationship to clinical ocular inflammatory disease. *J. Autoimmun.*, **9**, 575–585.
 78. O'DUFFY J. D., CALAMIA K., COHEN S., GORONZY J. J., HERMAN D., JORIZZO J., WEYAND C. and MATTESON E. (1998): Interferon- α treatment of Behcet's disease. *J. Rheumatol.*, **25**, 1938–1944.
 79. OKADA A. A., KEINO H., FUKAI T., SAKAI J., USUI M. and MIZUGUCHI J. (1998): Effect of type I interferon on experimental autoimmune uveoretinitis in rats. *Ocul. Immunol. Inflamm.*, **6**, 215–226.
 80. OKADA A. A., KEINO H., SUZUKI J., SAKAI J., USUI M. and MIZUGUCHI J. (1998): Kinetics of intraocular cytokines in the suppression of experimental autoimmune uveoretinitis by type I IFN. *Int. Immunol.*, **10**, 1917–1922.
 81. OKADA A. A., SAKAI J., USUI M. and MIZUGUCHI J. (1998): Intraocular cytokine quantification of experimental autoimmune uveoretinitis in rats. *Ocul. Immunol. Inflamm.*, **6**, 111–120.
 82. OPPMANN B., LESLEY R., BLOM B., TIMANS J. C., XU Y., HUNTE B., VEGA F., YU N., WANG J., SINGH K., ZONIN F., VAISBERG E., CHURAKOVA T., LIU M., GORMAN D., WAGNER J., ZURAWSKI S., LIU Y., ABRAMS J. S., MOORE K. W., RENNICK D., DE WAAL-MALEFYT R., HANNUM C., BAZAN J. F. and KASTELEIN R. A. (2000): Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity*, **13**, 715–725.
 83. OSHIMA K., YANASE N., IBUKIYAMA C., YAMASHINA A., KAYAGAKI N., YAGITA H. and MIZUGUCHI J. (2001): Involvement of TRAIL/TRAIL-R interaction in IFN- α -induced apoptosis of DAUDI b lymphoma cells. *Cytokine*, **14**, 193–201.
 84. PARRONCHI P., DE CARLI M., MANETTI R., SIMONELLI C., SAMPOGNARO S., PICCINNI M. P., MACCHIA D., MAGGI E., DEL PRETE G. and ROMAGNANI S. (1992): IL-4 and IFN (α and γ) exert opposite regulatory effects on the development of cytolytic potential by Th1 or Th2 human T cell clones. *J. Immunol.*, **149**, 2977–2983.
 85. POWELL M. B., MITCHELL D., LEDERMAN J., BUCKMEIER J., ZAMVIL S. S., GRAHAM M., RUDDLE N. H. and STEINMAN L. (1990): Lymphotoxin and tumor necrosis factor- α production by myelin basic protein-specific T cell clones correlates with encephalitogenicity. *Int. Immunol.*, **2**, 539–544.
 86. PRENDERGAST R. A., ILIFF C. E., COSKUNCAN N. M., CASPI R. R., SARTANI G., TARRANT T. K., LUTTY G. A. and MCLEOD D. S. (1998): T cell traffic and the inflammatory response in experimental autoimmune uveoretinitis. *Invest. Ophthalmol. Vis. Sci.*, **39**, 754–762.
 87. PULENDRAN B., PALUCKA K. and BANCHEREAU J. (2001): Sensing pathogens and tuning immune responses. *Science*, **293**, 253–256.
 88. RAMANATHAN S., DE KOZAK Y., SAOUDI A., GOUREAU O., VAN DER MEIDE P. H., DRUET P. and BELLON B. (1996): Recombinant IL-4 aggravates experimental autoimmune uveoretinitis in rats. *J. Immunol.*, **157**, 2209–2215.
 89. REIS E SOUSA C. (2001): Dendritic cells as sensors of infection. *Immunity*, **14**, 495–498.
 90. RIZZO L. V., XU H., CHAN C. C., WIGGERT B. and CASPI R. R. (1998): IL-10 has a protective role in experimental autoimmune uveoretinitis. *Int. Immunol.*, **10**, 807–814.
 91. SANTINI S. M., LAPENTA C., LOGOZZI M., PARLATO S., SPADA M., DI PUCCHIO T. and BELARDELLI F. (2000): Type I interferon as a powerful adjuvant for monocyte-derived dendritic cell development and activity *in vitro* and in Hu-PBL-SCID mice. *J. Exp. Med.*, **191**, 1777–1788.

92. SAOUDI A., KUHN J., HUYGEN K., DE KOZAK Y., VELU T., GOLDMAN M., DRUET P. and BELLON B. (1993): TH2 activated cells prevent experimental autoimmune uveoretinitis, a TH1-dependent autoimmune disease. *Eur. J. Immunol.*, **23**, 3096–3103.
93. SARAWAR S. R. and DOHERTY P. C. (1994): Concurrent production of interleukin-2, interleukin-10, and gamma interferon in the regional lymph nodes of mice with influenza pneumonia. *J. Virol.*, **68**, 3112–3119.
94. SHEVACH E. M., CHANG J. T. and SEGAL B. M. (1999): The critical role of IL-12 and the IL-12R beta 2 subunit in the generation of pathogenic autoreactive Th1 cells. *Springer Semin. Immunopathol.*, **21**, 249–262.
95. STARK G. R., KERR I. M., WILLIAMS B. R., SILVERMAN R. H. and SCHREIBER R. D. (1998): How cells respond to interferons. *Annu. Rev. Biochem.*, **67**, 227–264.
96. STEWART T. A., HULTGREN B., HUANG X., PITTS-MEEK S., HULLY J. and MACLACHLAN N. J. (1993): Induction of type I diabetes by interferon-alpha in transgenic mice. *Science*, **260**, 1942–1946.
97. STOCKINGER B. (1999): T lymphocyte tolerance: from thymic deletion to peripheral control mechanisms. *Adv. Immunol.*, **71**, 229–265.
98. STREILEIN J. W. (1987): Immune regulation and the eye: a dangerous compromise. *FASEB J.*, **1**, 199–208.
99. SU S. B., SILVER P. B., ZHANG M., CHAN C. C. and CASPI R. R. (2001): Pertussis toxin inhibits induction of tissue-specific autoimmune disease by disrupting G protein-coupled signals. *J. Immunol.*, **167**, 250–256.
100. SUEDA J., HIKITA N., MOCHIZUKI M., JIMI A. and KOJIRO M. (2000): Kinetics of apoptotic cells in experimental autoimmune uveoretinitis. *Invest. Ophthalmol. Vis. Sci.*, **41**, 799–804.
101. TAKEUCHI M., ALARD P. and STREILEIN J. W. (1998): TGF-beta promotes immune deviation by altering accessory signals of antigen-presenting cells. *J. Immunol.*, **160**, 1589–1597.
102. TAKEUCHI M., KEZUKA T., INOUE H., SAKAI J., USUI M., TAKAHASHI T. and TAGUCHI O. (1998): Suppression of spontaneous uveoretinitis development by non-immunopathogenic peptide immunization. *Eur. J. Immunol.*, **28**, 1578–1586.
103. TAKEUCHI M., KOSIEWICZ M. M., ALARD P. and STREILEIN J. W. (1997): On the mechanisms by which transforming growth factor-beta 2 alters antigen-presenting abilities of macrophages on T cell activation. *Eur. J. Immunol.*, **27**, 1648–1656.
104. TAKEUCHI M., YOKOI H., TSUKAHARA R., SAKAI J. and USUI M. (2001): Differentiation of Th1 and Th2 cells in lymph nodes and spleens of mice during experimental autoimmune uveoretinitis. *Jpn. J. Ophthalmol.*, **45**, 463–469.
105. TANAKA N., SATO M., LAMPHIER M. S., NOZAWA H., ODA E., NOGUCHI S., SCHREIBER R. D., TSUJIMOTO Y. and TANIGUCHI T. (1998): Type I interferons are essential mediators of apoptotic death in virally infected cells. *Genes Cells*, **3**, 29–37.
106. TARRANT T. K., SILVER P. B., CHAN C. C., WIGGERT B. and CASPI R. R. (1998): Endogenous IL-12 is required for induction and expression of experimental autoimmune uveitis. *J. Immunol.*, **161**, 122–127.
107. TARRANT T. K., SILVER P. B., WAHLSTEN J. L., RIZZO L. V., CHAN C. C., WIGGERT B. and CASPI R. R. (1999): Interleukin 12 protects from a T helper type 1-mediated autoimmune disease, experimental autoimmune uveitis, through a mechanism involving interferon gamma, nitric oxide, and apoptosis. *J. Exp. Med.*, **189**, 219–230.
108. TAYLOR A. W. (1996): Neuroimmunomodulation in immune privilege: role of neuropeptides in ocular immunosuppression. *Neuroimmunomodulation*, **3**, 195–204.
109. THILLAYE-GOLDENBERG B., GOUREAU O., NAUD M. C. and DE KOZAK Y. (2000): Delayed onset and decreased severity of experimental autoimmune uveoretinitis in mice lacking nitric oxide synthase type 2. *J. Neuroimmunol.*, **110**, 31–44.
110. TILG H. and PESCHEL C. (1996): Interferon-alpha and its effects on the cytokine cascade: a pro- and anti-inflammatory cytokine. *Leuk. Lymphoma*, **23**, 55–60.
111. TOMPKINS W. A. (1999): Immunomodulation and therapeutic effects of the oral use of interferon-alpha: mechanism of action. *J. Interferon Cytokine Res.*, **19**, 817–828.
112. TSAMBAOS D., EICHELBURG D. and GOOS M. (1986): Behcet's syndrome: treatment with recombinant leukocyte alpha-interferon. *Arch. Dermatol. Res.*, **278**, 335–336.
113. UCHIO E., KIJIMA M., TANAKA S. and OHNO S. (1994): Suppression of experimental uveitis with monoclonal antibodies to ICAM-1 and LFA-1. *Invest. Ophthalmol. Vis. Sci.*, **35**, 2626–2631.
114. UFRET-VINCENTY R. L., QUIGLEY L., TRESSER N., PAK S. H., GADO A., HAUSMANN S., WUCHERPFENNIG K. W. and BROCKE S. (1998): *In vivo* survival of viral antigen-specific T cells that induce experimental autoimmune encephalomyelitis. *J. Exp. Med.*, **188**, 1725–1738.
115. VIAL T. and DESCOTES J. (1995): Immune-mediated side-effects of cytokines in humans. *Toxicology*, **105**, 31–57.
116. WAHLSTEN J. L., GITCHELL H. L., CHAN C. C., WIGGERT B. and CASPI R. R. (2000): Fas and Fas ligand expressed on cells of the immune system, not on the target tissue, control induction of experimental autoimmune uveitis. *J. Immunol.*, **165**, 5480–5486.
117. WAKEFIELD D., CUELLO C., DI GIROLAMO N. and LLOYD A. (1999): The role of cytokines and chemokines in uveitis. *Dev. Ophthalmol.*, **31**, 53–66.
118. WALDNER H., SOBEL R. A., HOWARD E. and KUCHROO V. K. (1997): Fas- and FasL-deficient mice are resistant to induction of autoimmune encephalomyelitis. *J. Immunol.*, **159**, 3100–3103.
119. WANG X., CHEN M., WANDINGER K. P., WILLIAMS G. and DHIB-JALBUT S. (2000): IFN-beta-1b inhibits IL-12 production in peripheral blood mononuclear cells in an IL-10-dependent mechanism: relevance to IFN-beta-1b therapeutic effects in multiple sclerosis. *J. Immunol.*, **165**, 548–557.
120. WHITCUP S. M., DEBARGE L. R., CASPI R. R., HARNING R., NUSSENBLATT R. B. and CHAN C. C. (1993): Monoclonal antibodies against ICAM-1 (CD54) and LFA-1 (CD11a/CD18) inhibit experimental autoimmune uveitis. *Clin. Immunol. Immunopathol.*, **67**, 143–150.
121. WILBANKS G. A., MAMMOLENTI M. and STREILEIN J. W. (1991): Studies on the induction of anterior chamber-associated immune deviation (ACAID). II. Eye-derived cells participate in generating blood-borne signals that induce ACAID. *J. Immunol.*, **146**, 3018–3024.
122. XU H., RIZZO L. V., SILVER P. B. and CASPI R. R. (1997): Uveitogenicity is associated with a Th1-like lymphokine profile: cytokine-dependent modulation of early and committed effector T cells in experimental autoimmune uveitis. *Cell. Immunol.*, **178**, 69–78.

123. XU H., WAWROUSEK E. F., REDMOND T. M., NICKERSON J. M., WIGGERT B., CHAN C. C. and CASPI R. R. (2000): Transgenic expression of an immunologically privileged retinal antigen extraocularly enhances self tolerance and abrogates susceptibility to autoimmune uveitis. *Eur. J. Immunol.*, **30**, 272–278.
124. YOKOI H., KATO K., KEZUKA T., SAKAI J., USUI M., YAGITA H. and OKUMURA K. (1997): Prevention of experimental autoimmune uveoretinitis by monoclonal antibody to interleukin-12. *Eur. J. Immunol.*, **27**, 641–646.
125. ZHANG X., SUN S., HWANG I., TOUGH D. F. and SPRENT J. (1998): Potent and selective stimulation of memory-phenotype CD8⁺ T cells *in vivo* by IL-15. *Immunity*, **8**, 591–599.

Received in December 2001

Accepted in April 2002