

Transcriptional regulation in thymic epithelial cells for the establishment of self tolerance

Mitsuru Matsumoto

Division of Molecular Immunology, Institute for Enzyme Research, University of Tokushima, Tokushima, Japan

Received: 2006.10.10, **Accepted:** 2006.11.30, **Published online first:** 2007.02.02

Abstract

Thymic epithelial cells (TECs) play pivotal roles in the establishment of self tolerance through critical dialogue with developing thymocytes. Unique actions of two transcriptional regulators within TECs, NF- κ B-inducing kinase (NIK) and an autoimmune regulator (AIRE), for the establishment of self tolerance have recently been highlighted by studies using a strain of mouse bearing a natural mutation of the *NIK* gene (*aly* mice) and gene-targeted mice, respectively. Previous studies have demonstrated essential roles of NIK downstream of the lymphotoxin- β receptor (LT β R), which is essential for the development of secondary lymphoid organs; *aly* mice lack all lymph nodes and Peyer's patches because of the defective LT β R signaling. Additional roles of NIK in thymic organogenesis downstream of LT β R, mainly through the developmental regulation of TECs, have now emerged, although the corresponding ligand(s) for LT β R participating in this action have not been fully characterized. In contrast, *AIRE*, a gene responsible for the development of an organ-specific autoimmune disease that demonstrates monogenic autosomal recessive inheritance, contributes to the establishment of self tolerance probably by controlling the expression of self antigens through yet undetermined molecular mechanisms. Thus, it is highly likely that a group of genes control self tolerance within TECs through unique and coordinated actions, and that an understanding of this process would help to unravel the pathogenesis of autoimmune disease.

Key words: autoimmune disease, thymic epithelial cell, NF- κ B-inducing kinase, *aly* mice, lymphotoxin, AIRE.

Corresponding author: Mitsuru Matsumoto, M.D., Ph.D., Division of Molecular Immunology, Institute for Enzyme Research, University of Tokushima, 3-18-15 Kuramoto, Tokushima 770-8503, Japan, tel.: +81 88 633-74-32, fax: +81 88 633-74-34, e-mail: mitsuru@ier.tokushima-u.ac.jp

INTRODUCTION

Autoimmune disease is a pathological condition in which the immune system turns on itself and causes serious damage to host tissues through as yet unknown mechanisms [33]. Clarification of a unifying concept for the mechanisms underlying the development of autoimmune disease has been one of the major challenges of immunological studies. Breakdown of self tolerance is considered to be the key event in initiating the disease process, and an understanding the pathogenesis involved is crucial for developing a suitable therapeutic approach. For this reason, it is essential to know how self tolerance is established within the organized thymic microenvironment.

Physical contact between thymocytes and the thymic stroma is essential for T cell maturation and shapes the T cell repertoire in the periphery [39]. Stromal elements that control these processes still remain elusive. Recently, much attention has been paid to the epithelial-cell component of the stroma (i.e. thymic epithelial

cells – TECs), since increasing numbers of mutant and gene-targeted mice bearing structural and/or functional TEC defects, many of which are actually associated with autoimmune disease phenotypes, have been reported. In this brief review, I will focus on two transcriptional regulators within TECs, NF- κ B-inducing kinase (NIK) and the autoimmune regulator (AIRE), which play crucial roles in the establishment of self tolerance by maintaining the developmental and functional integrity of TECs, respectively, thereby preventing the development of autoimmune disease (Fig. 1).

NIK

NIK is structurally related to mitogen-activated protein kinase kinase kinase [43] and has been shown to phosphorylate both I κ B kinase (IKK)- α and IKK- β , which sequentially activate the downstream I κ B proteins necessary for NF- κ B activation [40]. The alympho-

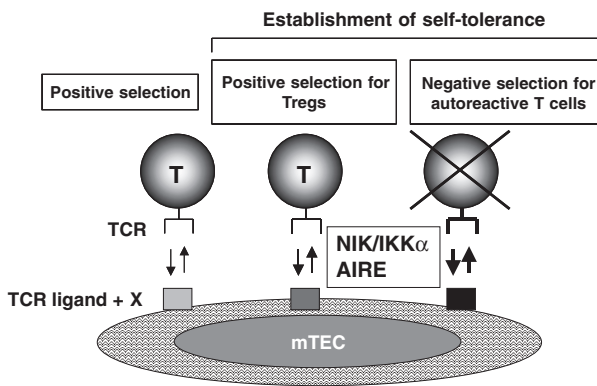


Fig. 1. The transcriptional regulators NIK/IKK α and AIRE within mTECs establish self tolerance through critical dialogues with developing thymocytes. Two known mechanisms of self tolerance, i.e. positive selection for Tregs and negative selection for autoreactive T cells, depending on the avidity of the interactions (depicted by the different depths of shading) between TCR and its self ligands expressed on mTECs are shown. "X" denotes costimulatory pathways and/or undetermined molecules that might additionally contribute to this process.

plasia (*aly*) strain of mouse carries a natural mutation of the *NIK* gene [47, 57] in which a G855R substitution in the C-terminus of the protein results in an inability to bind to IKK α [46] (Fig. 2). *aly* mice have provided a unique model for the abnormal development of lymphoid organs, since they lack all lymph nodes and Peyer's patches, and the development of spleen architectural features, such as germinal centers and follicular dendritic cell clusters, is disturbed [47, 57, 64]. This is due to defective NF- κ B activation through the lymphotoxin (LT)- β receptor (LT β R) [45, 46, 57], which is essential for the development of secondary lymphoid organs [44]. Thymic structure is also disorganized in *aly* mice; the medulla in *aly* mice is smaller than that in control mice, and the boundary of the cortex and medulla is unclear [32, 47, 48, 57]. Importantly, *aly* mice also serve as a model of autoimmune disease, but of unknown etiology [59]; histopathological analysis of *aly* mice has revealed chronic inflammatory changes in several organs, including the liver, pancreas, lung, salivary gland and lacrimal gland [32, 47, 59]. We reasoned that the autoimmune-disease phenotype seen in *aly* mice might be associated with the altered thymic microenvironment. This hypothesis was later proven correct, when thymic chimeras were generated [32]. *aly* mouse thymus-grafted nude mice showed marked lymphoid cell infiltration in the liver and pancreas, accompanied by autoantibody production. In contrast, control thymus-grafted nude mice showed no such changes. Histological evaluation of the grafted thymus revealed that control thymus contained cells reactive with the lectin *Ulex europaeus* agglutinin 1 (UEA-1)⁺, whereas *aly* mouse embryonic thymus grafted into nude mice did not acquire UEA-1⁺ cells, suggesting that production of UEA-1⁺ medullary epithelial cells requires normal NIK in the thymic stromal element.

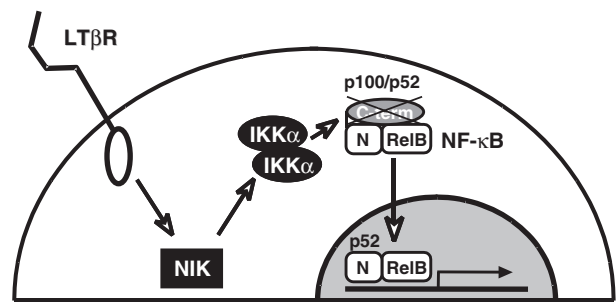


Fig. 2. Alternative NF- κ B activation pathway downstream of LT β R within TECs. LT β R ligation activates NIK by yet undetermined molecular mechanisms that result in the activation of IKK α homodimers. Activated IKK α catalyzes p100 of NF- κ B2 through ubiquitin-dependent degradation of the C-terminus and generates p52, a mature form of NF- κ B2 consisting of the N-terminus. p52 coupled with RelB enters the nucleus to activate the target genes necessary for the differentiation of TECs. *NIK*-mutation in *aly* mice results in inability to bind to and activate IKK α .

Although the exact mechanism by which NIK regulates the thymic microenvironment required for the establishment of central tolerance is unknown, the disorganized thymic structure together with reduced *Aire* expression in mice with a mutation disrupting the *RelB* gene merits attention [26]. Because of the phenotypic similarities between *aly* and *RelB*-deficient mice (multi-inflammatory lesions together with absence of UEA-1⁺ medullary epithelial cells in the thymus) [62], we speculate that NIK regulates the thymic microenvironment through activation of the NF- κ B complex containing RelB. As the production of p52 is impaired in *aly* mouse thymic stroma [32] and NIK has been shown to be necessary for the production of natural killer T cells through the action of RelB [18, 58], it is reasonable to speculate that the NIK-related signaling pathway(s) activates the NF- κ B complex in the thymic stroma consisting mainly of p52/RelB heterodimers to generate the thymic microenvironment (Fig. 2).

LT β R and its ligand

Although NIK is an essential component downstream of LT β R [46, 57] and *aly* mice show structural abnormality of the thymus [32, 47, 48, 57], the finding that LT β R-deficient mice have a disorganized thymic structure was somewhat surprising [9] because none of the preceding reports on single-gene knockout mice, either for the membrane-bound LT component (i.e. LT α and LT β) or LIGHT (two known ligands for LT β R [61]), referred to the thymic phenotypes in these mice [20, 44, 52]. In fact, deficiency of LT α alone was not accompanied by any obvious changes in thymic structure that would have resulted in reduced *Aire* expression at both the transcriptional [32] and protein levels (unpublished observation). LT β R-deficient mice show marked reduction of UEA-1⁺ cells caused by both loss of the characteristic 3-dimensional organization and a reduction in the absolute number of epithelial cells [9]. In

contrast, LT β -deficient mice show no significant reduction in the total mass of medullary TECs (mTECs), although changes in the UEA-1⁺ cell distribution pattern have been pointed out. Introduction of LIGHT deficiency in LT β -deficient mice resulted in no additional deterioration. Interestingly, LT β /LIGHT double-deficient mice (lack of both membrane-bound LT and LIGHT) showed less severe thymic disorganization than LT β R-deficient mice, suggesting that LT β R might have additional ligand(s) other than membrane-bound LT and LIGHT [9, 55]. Alternatively, in light of the fact that LT β -deficient mice show less profound phenotypes of lymph-node genesis (i.e. presence of mesenteric lymph nodes) compared with LT α -deficient mice [3, 36], it is possible that LT α /LIGHT double-deficient mice might show quite equivalent thymic phenotypes to those of LT β R-deficient mice. If this is the case, there may be a weak *in vivo* interaction between secreted LT (i.e. LT α homotrimer) and LT β R [55], although an *in vitro* study did not demonstrate such binding [12]. In either case, it would be important to note that deficiency in LT α (not LT β) or LT β R seems to result in different thymic phenotypes, even though both deficiencies produce very similar defects in secondary lymphoid organogenesis.

LT β R and autoimmunity

LT β R-deficient mice show some signs of autoimmunity; their serum contains autoantibodies against several organs (i.e. stomach, pancreas and salivary gland) [9]. However, it is not yet clear whether the perivascular accumulation of lymphocytes in multiple organs [20], which is also seen in mice deficient of LT α or LT β , represents an autoimmune status or an alteration of the lymphocyte distribution caused by defective secondary lymphoid organogenesis. How LT β R signaling controls the thymic microenvironment responsible for the establishment of central tolerance is debatable. One study suggested that LT β R ligation by membrane-LT regulates the transcription of tissue-restricted antigen (TRA) genes (for further discussion, see the section on AIRE) from established mTECs in an Aire-dependent manner [11], whereas another study concluded that LT β R affects mTEC differentiation, thereby controlling TRA gene transcription in the whole thymus, not from individual mTECs [9]. In the latter study, sorted mTECs from LT β R-deficient mice showed indistinguishable levels of both *Aire* and TRA gene expression when equal numbers of mTECs were examined [9]. Consistent with this, mTEC lines established from *aly* mice showed no reduction of TRA gene expression, although total thymus from *aly* mice showed much lower transcription levels of these genes [32, 35]. Thus, a role of LT β R in mTEC development, rather than transcriptional regulation of *Aire* (and TRA gene expression), seems to be a more likely explanation for the autoimmune phenotypes of LT β R-deficient mice. In this regard, it seems important to mention that the spectrum and/or features of autoimmunity are quite different between LT β R-

-deficient mice and Aire-deficient mice, mirrored by the difference in their autoimmune pathogenesis. An integrated and detailed phenotypic analysis of these mice should help to clarify many aspects of thymus biology.

LT β R signaling and more

NIK-IKK α constitutes an essential component downstream of LT β R for secondary lymphoid organogenesis [46]. It is therefore reasonable to speculate that NIK-IKK α also plays important roles in thymic organogenesis through the action of LT β R signaling. However, given that *aly* mice show more profound thymic reduction and disorganization of mTECs than LT β R-deficient mice [9], it is possible that in this process NIK-IKK α is additionally acting downstream on other receptor(s) beyond LT β R. One hint in the search for such receptor(s) involved in NIK-IKK α -dependent thymic organogenesis is impaired processing of NF- κ B2 in thymic stroma from *aly* mice [32] and IKK α -deficient mice [35]. This alternative NF- κ B activation pathway [56, 63] was originally demonstrated in hemopoietic cells from *aly* mice [64] and subsequently characterized for LT β R [13] (Fig. 2). Another signal that involves the generation of p52 from a precursor p100 might represent an additional NIK-IKK α -dependent pathway that could fill the gaps of thymic phenotypes between *aly* mice and LT β R-deficient mice.

NF- κ B activation within TEC

It is now clear that LT β R/NIK-IKK α is not the only NF- κ B-activating axis that regulates thymic organogenesis. Tumor necrosis factor receptor (TNFR)-associated factor 6 (TRAF6) has also been demonstrated to be essential for the organization of the thymic microenvironment [1]. TRAF6, an adapter molecule that transduces signals from members of the TNF superfamily and Toll/IL-1 receptor family, activates NF- κ B and activating protein 1 [29]. Similarly to *aly* mice and IKK α -deficient mice, TRAF6-deficient thymus-grafted nude mice show marked lymphoid cell infiltration in multiple organs. In contrast to *aly* mice and IKK α -deficient mice, however, NF- κ B2 processing in TECs from TRAF6-deficient mice is not impaired. Instead, RelB expression in TECs is severely reduced. TRAF6-dependent RelB expression has been confirmed by the recovery of RelB expression following the introduction of TRAF6 into TRAF6-deficient TECs. Thus, deficiency of NIK/IKK α and TRAF6 merges at the point where p52/RelB complex formation is disturbed, although this does not mean that NIK/IKK α and TRAF6 cooperate together within TECs in order for this heterodimeric complex to be formed. Rather, NIK/IKK α and TRAF6 probably regulate NF- κ B activation independently in this process, because TRAF6 deficiency does not affect NF- κ B activation downstream of LT β R [1]. The upstream receptor(s) responsible for TRAF6-dependent thymic organogenesis is currently unknown.

AIRE

Obviously, one of the critical roles of the thymic stroma in establishing self tolerance is the elimination of pathogenic autoreactive T cells by negative selection [27]. For this purpose, TECs need to express a set of self antigens (Ags) encompassing all the self Ags expressed by parenchymal organs, a phenomenon termed promiscuous gene expression (PGE) [15, 38]. Supporting this hypothesis, analysis of gene expression in the thymic stroma has demonstrated that epithelial cells of the medulla are a specialized cell type in which promiscuous expression of a broad range of TRA genes is an autonomous property [15, 39].

Negative selection by Aire

Because Aire-deficient mTECs show reduced transcription of TRA genes [6, 30] and Aire-deficient mice show impaired elimination of autoreactive T cells, as demonstrated with the use of transgenic mice expressing a neo-self Ag under the control of a tissue-specific promoter together with a T cell receptor (TCR) specific for the corresponding Ag [41, 42], it has been postulated that Aire regulates PGE to prevent autoimmunity; mice expressing hen egg lysozyme (HEL) in pancreatic β cells driven by the rat insulin promoter were crossed with mice expressing TCR specific for HEL, and the fate of HEL-specific T cells was monitored in either the presence or absence of Aire. Remarkably, Aire-deficient TCR-transgenic mice showed almost complete failure to delete the autoreactive (i.e. HEL-specific) T cells in the thymus [42]. Because Aire-deficient mTECs showed a reduction in transcription of a group of genes encoding peripheral Ags analyzed by the gene-chip technique [6], it has been hypothesized that pathogenic autoreactive T cells could not be eliminated efficiently due to the reduced expression of corresponding target Ags in the Aire-deficient thymus [41]. However, more recent studies have suggested that some other form of Aire-dependent negative selection is more plausible. Aire-deficient mice develop Sjögren's syndrome-like pathologic changes in the exocrine organs, and this is associated with autoimmunity against a ubiquitous protein, α -fodrin. Remarkably, transcriptional expression of α -fodrin is retained in the Aire-deficient thymus [37].

Because, unlike TRA, α -fodrin is a ubiquitously expressed actin-binding protein [25], one might argue that this finding may not be relevant for the assessment of the Aire-dependent negative selection process. However, subsequent studies using Aire-deficient NOD mice have demonstrated that the mice develop autoreactivity against a prototypic TRA whose transcription is not down-regulated in the thymus [50]. In Aire-deficient NOD mice, acinar cells rather than β -cell islets are the major targets of autoimmune destruction, and this alteration of intra-pancreatic target-organ specificity is associated with the production of autoantibody against pancreas-specific protein disulfide isomerase (PDIP), an Ag

expressed predominantly by acinar cells [16, 17]. Transcriptional expression of PDIP is retained in the Aire-deficient NOD thymus, further supporting the concept that Aire may regulate the survival of autoreactive T cells beyond the transcriptional control of self protein expression in the thymus [50]. Conversely, insulin is a prime target Ag recognized by autoreactive T cells in NOD mice [49], and TECs isolated from Aire-deficient NOD mice show reduced expression of insulin. Nevertheless, β -cell islets are relatively less affected by autoimmune attack in Aire-deficient NOD mice [50], which does not support the concept that Aire-dependent transcriptional control of TRA genes in TECs accounts for the development of Aire-dependent autoimmunity. Based on these findings, we suggest that Aire may regulate the processing and/or presentation of self Ags by TECs, possibly through coordinated action with bone marrow-derived cells [21], so that the maturing T cells can recognize the corresponding self Ags in a form capable of efficiently triggering autoreactive T cells. This alternative view of the function of Aire in the establishment of central tolerance has recently been supported by a study with transgenic mouse models [5]. In addition to these models, other Aire-dependent central-tolerance mechanisms as well as self tolerance in the periphery are also possible [53].

Aire in mTEC

Although it is already evident that loss of function of *AIRE* in humans and *Aire* in mice results in the breakdown of self tolerance [51], the physiological roles of *AIRE*/*Aire* that implicate self tolerance at the molecular level remain unresolved [2, 24, 60]. From a mechanistic viewpoint, I would suggest two possible models in which Aire exerts its function (Fig. 3). First, Aire plays

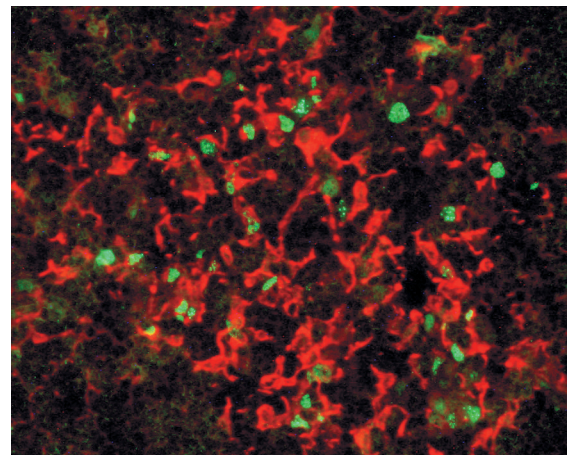


Fig. 3. Aire-positive cells in the thymus. Immunohistochemical staining using antibody against mouse Aire demonstrates Aire-expressing cells (stained in green) scattered in the medullary region of the thymus (stained in red with anti-ER-TR5 monoclonal antibody). Note that Aire is present within mTECs as nuclear dots of protein.

a tolerogenic role within particular type(s) of mTECs, which can be discerned as Aire-expressing cells when Aire is present (model 1). In other words, the presence of Aire within the cells is necessary in order for them to function normally as tolerance-establishing cells. In this scenario, Aire-deficient thymus may still contain such cell type(s) but without any tolerogenic function because they lack Aire within them. The second model hypothesizes that Aire is necessary for the development of particular cell type(s), including Aire-positive cells themselves, with tolerogenic function in the thymus (model 2). In this case, we assume that what are called Aire-positive cells and other Aire-dependent cell type(s) do not develop in the absence of Aire. In order to discriminate these possibilities, we need to establish specific markers of cells other than those with Aire expression that would make it possible to monitor the developmental process of Aire-bearing cells. If the former model is correct, then Aire-deficiency would not affect the development of phantom Aire-positive cells. In contrast, the latter model would reveal any lack of Aire-dependent cell type(s), again including so-called Aire-positive cells, in the absence of Aire, although extensive immunohistochemical analysis has so far not demonstrated any obvious changes in Aire-deficient thymus [37, 50].

Although both models define Aire-positive cells as a unique cell type among mTECs, we still do not know whether Aire-positive cells are the predominant cell type(s) responsible for the induction of self tolerance. Rather, Aire-positive cells may regulate the function of other tolerogenic mTECs. Similarly, it is not yet clear whether Aire-positive cells are the major source of PGE from mTECs. In fact, a recent single-cell analysis has demonstrated that the expression of Aire in a mTEC is not sufficient for simultaneous co-expression of Aire-dependent TRA genes [23]. Establishment of specific markers for Aire-positive cells may help to clarify these issues as well.

Terminal differentiation model vs. developmental model

One of the key issues for acquiring an understanding of central tolerance is how we accept the dogma that the thymus should and does express a variety of self Ags for the elimination of self-reactive T cells. There are two contrasting models of how mTECs acquire their unique ability to express a broad range of self Ags (i.e. PGE) [22]. The terminal differentiation model assumes that mTECs eventually gain properties of PGE by becoming differentiated, more mature, and more promiscuous [14]. This model suggests that mTECs, especially Aire-positive cells, are specialized cell type(s) that have acquired this property by differentiation. In contrast, the developmental model considers that PGE is a reflection of the multi-potency of immature mTECs before the determination of developmental fate into particular cell type(s) [22]. In this model, the expression of a broad spectrum of TRA genes is considered to be regulated by

conserved developmental programs active in developing mTECs. These two models also pose different viewpoints regarding the roles of AIRE/Aire. The terminal differentiation model assumes that AIRE/Aire acts as an important player for the PGE within differentiated mTECs. This is based on the observation that PGE is correlated with CD80 expression levels on mTECs, and that CD80^{high} mTECs from Aire-deficient mice show a reduction in both the total number of overexpressed genes and the relative percentage of TRA genes [14]. In contrast, the developmental model considers that Aire is an important differentiation factor that determines the fate of immature mTECs [22]. As a result, Aire expression does not have to guarantee PGE, as described above [23]. Although not exclusively, the terminal differentiation model and developmental model may respectively favor models 1 and 2 proposed above for the roles of AIRE/Aire.

Regulatory T cells

In addition to negative selection, self tolerance is maintained by another mechanism involving immunoregulatory T cells (Treg) [54], and *Foxp3*, a transcription factor that is genetically defective in an autoimmune disease known as IPEX (immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome), is a key regulator for the development of Tregs [19, 28, 34]. So far, no obvious changes in the numbers [6, 37, 42, 50] or function [5, 37] of Tregs have been described in Aire-deficient mice. As a result, Aire/*Foxp3* double-deficient mice show fulminant autoimmunity in very early life and a gravely shortened life-span in comparison with mice that are defective in either gene alone [10]. Given that Tregs arise from relatively high-avidity interactions with self-peptide-MHC complexes just below the threshold for negative selection [7, 31, 54], it is somewhat unexpected that Aire deficiency has no major impact on the production or suppressive function of Tregs in both non-autoimmune-prone mice [5, 37] and autoimmune-prone NOD mice [50]. In contrast, *aly* mice [32] and TRAF6-deficient mice [1], both of which show abnormal development of TECs, have reduced numbers of Tregs. The differential requirement of NIK/TRAF6 and Aire for the production of Tregs but the need for all three factors for negative selection, even though impaired negative selection in NIK-mutant mice and TRAF6-deficient mice has not yet been fully demonstrated, might provide a clue as to how the negative selection and production of Tregs are divided, despite the fact that both decisions are dependent on common interactions with TECs, as illustrated in Fig. 1.

CONCLUSIONS

Table 1 shows a general phenotypic comparison between NIK-mutant *aly* mice together with IKK α -deficient mice, and Aire-deficient mice. Even for the two stro-

Table 1. Autoimmune pathogenesis in mice deficient of transcriptional regulators in TECs

	NIK-mutant mice IKK α -deficient mice	Aire-deficient mice
Thymic structure assessed by immu- nohistochemistry	Defective develop- ment of mTECs	Grossly normal
Thymic TRA gene expression detected by RT-PCR	Dramatically redu- ced in total thymus, but not in individual TECs	Reduced on an individual TEC basis, but not for all TRA genes
Production and function of Tregs	Reduced, but func- tionally competent	Unchanged
Negative selection studied with trans- genic mice	Not assessed, but probably defective	Defective
Possible mecha- nisms for the breakdown of central tolerance	Defective NF- κ B activation required for the development of mTECs	Defective pro- cessing and/or presentation of self Ag

mal factors referred to in this article, the exact mechanisms by which they contribute to the establishment of self tolerance still remain unresolved. Although much attention has been paid to the transcriptional regulation of self-Ag expression in mTECs, the precise extent to which its fluctuation might affect the processes of central tolerance per se are still unclear [4]. In this regard, it would also be important to bear in mind that assessment of negative selection using TCR-transgenic mice crossed with transgenic mice overexpressing the corresponding neo-self Ags may not reflect the exact physiological process that contributes to the prevention of autoimmunity [8]. Finally, a more fundamental and persistent issue related to self tolerance is what constitutes the “immunological self” for the immune system. There may be other means by which the thymic stroma achieves expression of the immunological self beyond what I have conceptually outlined in the present article. If this is the case, then the other ways in which T cells achieve recognition of the immunological self might govern the self-non-self discrimination process by as yet additional and unknown rules, similar to the discovery of microorganism recognition by Toll-like receptors, which opened a new era of immunological research.

Acknowledgment: This work was supported by Special Coordination Funds of the Ministry of Education, Culture, Sports, Science, and Technology (MEXT), and by Grant-in-Aid for Scientific Research from the MEXT (17047028, 17390291 and 18060030).

REFERENCES

1. Akiyama T., Maeda S., Yamane S., Ogino K., Kasai M., Kajiura F., Matsumoto M. and Inoue J. (2005): Dependence of self-tolerance on TRAF6-directed development of thymic stroma. *Science*, **308**, 248–251.
2. Akiyoshi H., Hatakeyama S., Pitkänen J., Mouri Y., Doucas V., Kudoh J., Tsurugaya K., Uchida D., Matsushima A., Oshikawa K., Nakayama K. I., Shimizu N., Peterson P. and Matsumoto M. (2004): Subcellular expression of autoimmune regulator (AIRE) is organized in a spatiotemporal manner. *J. Biol. Chem.*, **279**, 33984–33991.
3. Alimzhanov M. B., Kuprash D. V., Kosco-Vilbois M. H., Luz A., Turetskaya R. L., Tarakhovsky A., Rajewsky K., Nedospasov S. A. and Pfeffer K. (1997): Abnormal development of secondary lymphoid tissues in lymphotoxin β -deficient mice. *Proc. Natl. Acad. Sci. USA*, **94**, 9302–9307.
4. Anderson A. C. and Kuchroo V. K. (2003): Expression of self-antigen in the thymus: a little goes a long way. *J. Exp. Med.*, **198**, 1627–1629.
5. Anderson M. S., Venanzi E. S., Chen Z., Berzins S. P., Benoist C. and Mathis D. (2005): The cellular mechanism of Aire control of T cell tolerance. *Immunity*, **23**, 227–239.
6. Anderson M. S., Venanzi E. S., Klein L., Chen Z., Berzins S. P., Turley S. J., von Boehmer H., Bronson R., Dierich A., Benoist C. and Mathis D. (2002): Projection of an immunological self shadow within the thymus by the aire protein. *Science*, **298**, 1395–1401.
7. Apostolou I., Sarukhan A., Klein L. and von Boehmer H. (2002): Origin of regulatory T cells with known specificity for antigen. *Nat. Immunol.*, **3**, 756–763.
8. Baldwin T. A., Sandau M. M., Jameson S. C. and Hogquist K. A. (2005): The timing of TCR α expression critically influences T cell development and selection. *J. Exp. Med.*, **202**, 111–121.
9. Boehm T., Scheu S., Pfeffer K. and Bleul C. C. (2003): Thymic medullary epithelial cell differentiation, thymocyte emigration, and the control of autoimmunity require lympho-epithelial cross talk via LT β R. *J. Exp. Med.*, **198**, 757–769.
10. Chen Z., Benoist C. and Mathis D. (2005): How defects in central tolerance impinge on a deficiency in regulatory T cells. *Proc. Natl. Acad. Sci. USA*, **102**, 14735–14740.
11. Chin R. K., Lo J. C., Kim O., Blink S. E., Christiansen P. A., Peterson P., Wang Y., Ware C. and Fu Y. X. (2003): Lymphotoxin pathway directs thymic Aire expression. *Nat. Immunol.*, **4**, 1121–1127.
12. Crowe P. D., VanArsdale T. L., Walter B. N., Ware C. F., Hession C., Ehrenfels B., Browning J. L., Din W. S., Goodwin R. G. and Smith C. A. (1994): A lymphotoxin- β -specific receptor. *Science*, **264**, 707–710.
13. Dejardin E., Droin N. M., Delhase M., Haas E., Cao Y., Makris C., Li Z. W., Karin M., Ware C. F. and Green D. R. (2002): The lymphotoxin- β receptor induces different patterns of gene expression via two NF- κ B pathways. *Immunity*, **17**, 525–535.
14. Derbinski J., Gabler J., Brors B., Tierling S., Jonnakuty S., Hergenhausen M., Peltonen L., Walter J. and Kyewski B. (2005): Promiscuous gene expression in thymic epithelial cells is regulated at multiple levels. *J. Exp. Med.*, **202**, 33–45.
15. Derbinski J., Schulte A., Kyewski B. and Klein L. (2001): Promiscuous gene expression in medullary thymic epithelial cells mirrors the peripheral self. *Nat. Immunol.*, **2**, 1032–1039.
16. Desilva M. G., Lu J., Donadel G., Modi W. S., Xie H., Notkins A. L. and Lan M. S. (1996): Characterization and chromosomal localization of a new protein disulfide isomerase, PDIP, highly expressed in human pancreas. *DNA Cell Biol.*, **15**, 9–16.

17. Desilva M. G., Notkins A. L. and Lan M. S. (1997): Molecular characterization of a pancreas-specific protein disulfide isomerase, PDIP. *DNA Cell Biol.*, **16**, 269–274.
18. Elewaut D., Shaikh R. B., Hammond K. J., De Winter H., Leishman A. J., Sidobre S., Turovskaya O., Prigozy T. I., Ma L., Banks T. A., Lo D., Ware C. F., Cheroutre H. and Kronenberg M. (2003): NIK-dependent RelB activation defines a unique signaling pathway for the development of $V\alpha 14i$ NKT cells. *J. Exp. Med.*, **197**, 1623–1633.
19. Fontenot J. D., Gavin M. A. and Rudensky A. Y. (2003): Foxp3 programs the development and function of $CD4^+CD25^+$ regulatory T cells. *Nat. Immunol.*, **4**, 330–336.
20. Fütterer A., Mink K., Luz A., Kosco-Vilbois M. H. and Pfeffer K. (1998): The lymphotoxin β receptor controls organogenesis and affinity maturation in peripheral lymphoid tissues. *Immunity*, **9**, 59–70.
21. Gallegos A. M. and Bevan M. J. (2004): Central tolerance to tissue-specific antigens mediated by direct and indirect antigen presentation. *J. Exp. Med.*, **200**, 1039–1049.
22. Gillard G. O. and Farr A. G. (2005): Contrasting models of promiscuous gene expression by thymic epithelium. *J. Exp. Med.*, **202**, 15–19.
23. Gillard G. O. and Farr A. G. (2006): Features of medullary thymic epithelium implicate postnatal development in maintaining epithelial heterogeneity and tissue-restricted antigen expression. *J. Immunol.*, **176**, 5815–5824.
24. Gozani O., Karuman P., Jones D. R., Ivanov D., Cha J., Lugovskoy A. A., Baird C. L., Zhu H., Field S. J., Lessnick S. L., Villasenor J., Mehrotra B., Chen J., Rao V. R., Brugge J. S., Ferguson C. G., Payastre B., Myszkowski D. G., Cantley L. C., Wagner G., Divecha N., Prestwich G. D. and Yuan J. (2003): The PHD finger of the chromatin-associated protein ING2 functions as a nuclear phosphoinositide receptor. *Cell*, **114**, 99–111.
25. Haneji N., Nakamura T., Takio K., Yanagi K., Higashiyama H., Saito I., Noji S., Sugino H. and Hayashi Y. (1997): Identification of α -fodrin as a candidate autoantigen in primary Sjögren's syndrome. *Science*, **276**, 604–607.
26. Heino M., Peterson P., Sillanpää N., Guerin S., Wu L., Anderson G., Scott H. S., Antonarakis S. E., Kudoh J., Shimizu N., Jenkinson E. J., Naquet P. and Krohn K. J. (2000): RNA and protein expression of the murine autoimmune regulator gene (Aire) in normal, RelB-deficient and in NOD mouse. *Eur. J. Immunol.*, **30**, 1884–1893.
27. Hogquist K. A., Baldwin T. A. and Jameson S. C. (2005): Central tolerance: learning self-control in the thymus. *Nat. Rev. Immunol.*, **5**, 772–782.
28. Hori S., Nomura T. and Sakaguchi S. (2003): Control of regulatory T cell development by the transcription factor FOXP3. *Science*, **299**, 1057–1061.
29. Inoue J., Ishida T., Tsukamoto N., Kobayashi N., Naito A., Azuma S. and Yamamoto T. (2000): Tumor necrosis factor receptor-associated factor (TRAF) family: adapter proteins that mediate cytokine signaling. *Exp. Cell Res.*, **254**, 14–24.
30. Johnnidis J. B., Venzani E. S., Taxman D. J., Ting J. P., Benoist C. O. and Mathis D. J. (2005): Chromosomal clustering of genes controlled by the aire transcription factor. *Proc. Natl. Acad. Sci. USA*, **102**, 7233–7238.
31. Jordan M. S., Boesteanu A., Reed A. J., Petrone A. L., Hohenbeck A. E., Lerman M. A., Naji A. and Caton A. J. (2001): Thymic selection of $CD4^+CD25^+$ regulatory T cells induced by an agonist self-peptide. *Nat. Immunol.*, **2**, 301–306.
32. Kajiura F., Sun S., Nomura T., Izumi K., Ueno T., Bando Y., Kuroda N., Han H., Li Y., Matsushima A., Takahama Y., Sakaguchi S., Mitani T. and Matsumoto M. (2004): NF- κ B-inducing kinase establishes self-tolerance in a thymic stroma-dependent manner. *J. Immunol.*, **172**, 2067–2075.
33. Kamradt T. and Mitchison N. A. (2001): Tolerance and autoimmunity. *N. Engl. J. Med.*, **344**, 655–664.
34. Khattri R., Cox T., Yasayko S. A. and Ramsdell F. (2003): An essential role for Scurfin in $CD4^+CD25^+$ T regulatory cells. *Nat. Immunol.*, **4**, 337–342.
35. Kinoshita D., Hirota F., Kaisho T., Kasai M., Izumi K., Bando Y., Mouri Y., Matsushima A., Niki S., Han H., Oshikawa K., Kuroda N., Maegawa M., Irahara M., Takeda K., Akira S. and Matsumoto M. (2006): Essential role of I κ B kinase α in thymic organogenesis required for the establishment of self-tolerance. *J. Immunol.*, **176**, 3995–4002.
36. Koni P. A., Sacca R., Lawton P., Browning J. L., Ruddle N. H. and Flavell R. A. (1997): Distinct roles in lymphoid organogenesis for lymphotoxins α and β revealed in lymphotoxin β -deficient mice. *Immunity*, **6**, 491–500.
37. Kuroda N., Mitani T., Takeda N., Ishimaru N., Arakaki R., Hayashi Y., Bando Y., Izumi K., Takahashi T., Nomura T., Sakaguchi S., Ueno T., Takahama Y., Uchida D., Sun S., Kajiura F., Mouri Y., Han H., Matsushima A., Yamada G. and Matsumoto M. (2005): Development of autoimmunity against transcriptionally unrepressed target antigen in the thymus of Aire-deficient mice. *J. Immunol.*, **174**, 1862–1870.
38. Kyewski B., Derbinski J., Gotter J. and Klein L. (2002): Promiscuous gene expression and central T-cell tolerance: more than meets the eye. *Trends Immunol.*, **23**, 364–371.
39. Kyewski B. and Klein L. (2006): A central role for central tolerance. *Annu. Rev. Immunol.*, **24**, 571–606.
40. Ling L., Cao Z. and Goeddel D. V. (1998): NF- κ B-inducing kinase activates IKK- α by phosphorylation of Ser-176. *Proc. Natl. Acad. Sci. USA*, **95**, 3792–3797.
41. Liston A., Gray D. H., Lesage S., Fletcher A. L., Wilson J., Webster K. E., Scott H. S., Boyd R. L., Peltonen L. and Goodnow C. C. (2004): Gene dosage – limiting role of Aire in thymic expression, clonal deletion, and organ-specific autoimmunity. *J. Exp. Med.*, **200**, 1015–1026.
42. Liston A., Lesage S., Wilson J., Peltonen L. and Goodnow C. C. (2003): Aire regulates negative selection of organ-specific T cells. *Nat. Immunol.*, **4**, 350–354.
43. Malinin N. L., Boldin M. P., Kovalenko A. V. and Wallach D. (1997): MAP3K-related kinase involved in NF- κ B induction by TNF, CD95 and IL-1. *Nature*, **385**, 540–544.
44. Matsumoto M., Fu Y. X., Molina H. and Chaplin D. D. (1997): Lymphotoxin- α -deficient and TNF receptor-I-deficient mice define developmental and functional characteristics of germinal centers. *Immunol. Rev.*, **156**, 137–144.
45. Matsumoto M., Iwamasa K., Rennert P. D., Yamada T., Suzuki R., Matsushima A., Okabe M., Fujita S. and Yokoyama M. (1999): Involvement of distinct cellular compartments in the abnormal lymphoid organogenesis in lymphotoxin- α -deficient mice and alymphoplasia (*aly*) mice defined by the chimeric analysis. *J. Immunol.*, **163**, 1584–1591.
46. Matsushima A., Kaisho T., Rennert P. D., Nakano H., Kurosawa K., Uchida D., Takeda K., Akira S. and Matsumoto M. (2001): Essential role of nuclear factor (NF)- κ B-inducing kinase and inhibitor of κ B (I κ B) kinase α in NF- κ B activation through lymphotoxin β receptor, but

- not through tumor necrosis factor receptor I. *J. Exp. Med.*, **193**, 631–636.
47. Miyawaki S., Nakamura Y., Suzuka H., Koba M., Yasumizu R., Ikehara S. and Shibata Y. (1994): A new mutation, *aly*, that induces a generalized lack of lymph nodes accompanied by immunodeficiency in mice. *Eur. J. Immunol.*, **24**, 429–434.
 48. Nakagawa K., Iwabuchi K., Ogasawara K., Ato M., Kajiwar M., Nishihori H., Iwabuchi C., Ishikura H., Good R. A. and Onoe K. (1997): Generation of NK1.1⁺ T cell antigen receptor α/β ⁺ thymocytes associated with intact thymic structure. *Proc. Natl. Acad. Sci. USA*, **94**, 2472–2477.
 49. Nakayama M., Abiru N., Moriyama H., Babaya N., Liu E., Miao D., Yu L., Wegmann D. R., Hutton J. C., Elliott J. F. and Eisenbarth G. S. (2005): Prime role for an insulin epitope in the development of type 1 diabetes in NOD mice. *Nature*, **435**, 220–223.
 50. Niki S., Oshikawa K., Mouri Y., Hirota F., Matsushima A., Yano M., Han H., Bando Y., Izumi K., Matsumoto M., Nakayama K. I., Kuroda N. and Matsumoto M. (2006): Alteration of intra-pancreatic target-organ specificity by abrogation of Aire in NOD mice. *J. Clin. Invest.*, **116**, 1292–1301.
 51. Peterson P. and Peltonen L. (2005): Autoimmune polyendocrinopathy syndrome type 1 (APS1) and AIRE gene: new views on molecular basis of autoimmunity. *J. Autoimmun.*, **25** (suppl.), 49–55.
 52. Pfeffer K. (2003): Biological functions of tumor necrosis factor cytokines and their receptors. *Cytokine Growth Factor Rev.*, **14**, 185–191.
 53. Ramsey C., Hassler S., Marits P., Kampe O., Surh C. D., Peltonen L. and Winqvist O. (2006): Increased antigen presenting cell-mediated T cell activation in mice and patients without the autoimmune regulator. *Eur. J. Immunol.*, **36**, 305–317.
 54. Sakaguchi S. (2004): Naturally arising CD4⁺ regulatory T cells for immunologic self-tolerance and negative control of immune responses. *Annu. Rev. Immunol.*, **22**, 531–562.
 55. Scheu S., Alferink J., Potzel T., Barchet W., Kalinke U. and Pfeffer K. (2002): Targeted disruption of LIGHT causes defects in costimulatory T cell activation and reveals cooperation with lymphotoxin β in mesenteric lymph node genesis. *J. Exp. Med.*, **195**, 1613–1624.
 56. Senftleben U., Cao Y., Xiao G., Greten F. R., Krahn G., Bonizzi G., Chen Y., Hu Y., Fong A., Sun S. C. and Karin M. (2001): Activation by IKK α of a second, evolutionary conserved, NF- κ B signaling pathway. *Science*, **293**, 1495–1499.
 57. Shinkura R., Kitada K., Matsuda F., Tashiro K., Ikuta K., Suzuki M., Kogishi K., Serikawa T. and Honjo T. (1999): A lymphoplasia is caused by a point mutation in the mouse gene encoding NF- κ B-inducing kinase. *Nat. Genet.*, **22**, 74–77.
 58. Sivakumar V., Hammond K. J., Howells N., Pfeffer K. and Weih F. (2003): Differential requirement for Rel/nuclear factor κ B family members in natural killer T cell development. *J. Exp. Med.*, **197**, 1613–1621.
 59. Tsubata R., Tsubata T., Hiai H., Shinkura R., Matsumura R., Sumida T., Miyawaki S., Ishida H., Kumagai S., Nakao K. and Honjo T. (1996): Autoimmune disease of exocrine organs in immunodeficient alymphoplasia mice: a spontaneous model for Sjögren's syndrome. *Eur. J. Immunol.*, **26**, 2742–2748.
 60. Uchida D., Hatakeyama S., Matsushima A., Han H., Ishido S., Hotta H., Kudoh J., Shimizu N., Doucas V., Nakayama K. I., Kuroda N. and Matsumoto M. (2004): AIRE functions as an E3 ubiquitin ligase. *J. Exp. Med.*, **199**, 167–172.
 61. Ware C. F. (2005): Network communications: lymphotoxins, LIGHT, and TNF. *Annu. Rev. Immunol.*, **23**, 787–819.
 62. Weih F., Carrasco D., Durham S. K., Barton D. S., Rizzo C. A., Ryseck R. P., Lira S. A. and Bravo R. (1995): Multiorgan inflammation and hematopoietic abnormalities in mice with a targeted disruption of RelB, a member of the NF- κ B/Rel family. *Cell*, **80**, 331–340.
 63. Xiao G., Harhaj E. W. and Sun S. C. (2001): NF- κ B-inducing kinase regulates the processing of NF- κ B2 p100. *Mol. Cell*, **7**, 401–409.
 64. Yamada T., Mitani T., Yorita K., Uchida D., Matsushima A., Iwamasa K., Fujita S. and Matsumoto M. (2000): Abnormal immune function of hemopoietic cells from alymphoplasia (*aly*) mice, a natural strain with mutant NF- κ B-inducing kinase. *J. Immunol.*, **165**, 804–812.